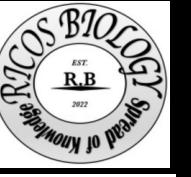




RICOS BIOLOGY JOURNAL



Vol. 4, no. 1
January, 2026



<https://ricosbiology.net>
<https://ricospublisher.us>
info@ricosbiology.net

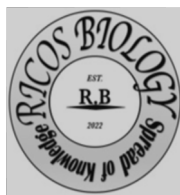


Table of contents

No.	Article	Pages
1	The Messenger RNA (mRNA) Revolution: From Fundamental Biology to Therapeutic Applications and Future Horizons. DOI: https://doi.org/10.33687/ricosbiol.04.01.106	1 - 11
2	The Epidemiological Nexus: Age, Geography, and Infectious Disease in Egyptian Cattle. DOI: https://doi.org/10.33687/ricosbiol.04.01.105	12 - 17
3	The Shape-Shifting Scourge: A Century of H1N1 Influenza - From Pandemic Terror to Persistent Threat. DOI: https://doi.org/10.33687/ricosbiol.04.01.102	18 - 35

The Messenger RNA (mRNA) Revolution: From Fundamental Biology to Therapeutic Applications and Future Horizons

Abouelhag H. A. *

*Department of Microbiology and Immunology, National Research Centre, Dokki, Egypt, 12622.

Received: 17-11-2025

Accepted: 20-01-2025

Published online: 30-01-2026

DOI: <https://doi.org/10.33687/ricosbiol.04.01.106>

Abstract

Messenger RNA (mRNA) has evolved from a fundamental biological intermediary to a versatile platform for therapeutic and prophylactic interventions. This review provides a comprehensive analysis of the mRNA field, beginning with the essential biology of natural mRNA processing and regulation. We detail the key engineering breakthroughs that transformed synthetic mRNA into a viable drug modality, including nucleoside modifications and sequence optimization to enhance stability and translational efficiency while modulating immunogenicity (Karikó, Buckstein, Ni, & Weissman, 2005; Pardi, Hogan, Porter, & Weissman, 2018). A critical discussion of delivery technologies, with a focus on lipid nanoparticles (LNPs), explains how these carriers enable *in vivo* application (Hou, Zaks, Langer, & Dong, 2021). The review then surveys the expansive therapeutic landscape, from the paradigm-shifting success of COVID-19 vaccines (Polack et al., 2020) to applications in protein replacement therapy, cancer immunotherapy, and gene editing. Finally, we examine persistent challenges—including delivery refinement, durability of response, and scaling manufacturing and envision future directions such as circular RNA, personalized neoantigen vaccines, and programmable protein therapeutics. The convergence of mRNA biology, chemistry, and delivery science heralds a new era in medicine with the potential to address a vast array of human diseases.

Keywords: mRNA, synthetic mRNA, mRNA therapeutics, lipid nanoparticles (LNPs), vaccinology, *in vitro* transcribed (IVT) mRNA, epitranscriptomics, RNA delivery, personalized medicine.

Introduction

1. The Central Dogma and the Historical Discovery of mRNA

The "Central Dogma" of molecular biology, articulated by Francis Crick, posits the unidirectional flow of genetic information from DNA to RNA to protein. The discovery of messenger RNA (mRNA) as the crucial intermediary in this pathway was a landmark achievement. In 1961, Sydney Brenner, François Jacob, and Matthew Meselson, through experiments on bacteriophage-infected *E. coli*, identified an unstable RNA fraction that carried genetic information from DNA to the ribosomes for protein synthesis (Brenner, Jacob, & Meselson, 1961). This ephemeral molecule, later termed messenger RNA, was characterized by its base sequence complementary to DNA and its rapid turnover, allowing cells to dynamically adjust their proteome in response to stimuli.

2. The Conceptual Leap: mRNA as a Therapeutic Platform

For decades, mRNA was studied primarily as a target for understanding gene regulation. The visionary idea of using synthetic mRNA as a drug emerged in the 1990s. Pioneering work by scientists like Jon Wolff demonstrated that *in vitro* transcribed (IVT) mRNA could be delivered to cells and animals to produce a functional protein (Wolff et al., 1990). However, major hurdles—namely, intrinsic immunogenicity triggering inflammatory responses, rapid enzymatic degradation, and inefficient *in vivo* delivery—stymied progress. The transformative breakthrough came with the discovery that incorporating modified nucleosides (e.g., pseudouridine) into IVT mRNA dramatically reduced its recognition by pattern recognition receptors, suppressing unwanted interferon responses and enhancing protein production (Karikó et al., 2005). This, coupled with advances in nanocarrier delivery, propelled mRNA from a laboratory tool to a clinical reality.

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



3. The COVID-19 Catalyst and Beyond

The SARS-CoV-2 pandemic served as an unprecedented validation and accelerator for mRNA technology. The rapid development, stunning efficacy, and global deployment of mRNA-based COVID-19 vaccines (mRNA-1273 and BNT162b2) demonstrated the platform's key advantages: speed (design based on sequence alone), flexibility (easy targeting of new variants), potency (strong humoral and cellular immunity), and scalable manufacturing (Polack et al., 2020; Corbett et al., 2020). This success has unleashed vast investment and interest, expanding the therapeutic horizon far beyond infectious diseases.

4. Scope and Aims of This Review

This article aims to provide a holistic overview of the mRNA revolution. We will first elucidate the biology of natural mRNA to establish a foundational understanding. We will then dissect the engineering principles behind synthetic mRNA and the delivery technologies that make it functional *in vivo*. A comprehensive survey of current and emerging therapeutic applications will follow. Finally, we will confront the remaining challenges and outline future research directions that will define the next decade of mRNA-based medicine.

The Biology of Natural mRNA: Structure, Processing, and Function

1. Canonical Structure and Function of mRNA Elements

A mature eukaryotic mRNA is a complex ribonucleoprotein particle with distinct functional regions:

- i. 5' Cap (7-methylguanosine): Protects from 5' exonucleases, facilitates ribosome binding during translation initiation, and is involved in splicing and nuclear export.
- ii. 5' Untranslated Region (UTR): Contains regulatory elements that control translation efficiency, stability, and subcellular localization. Secondary structures in the 5' UTR can influence ribosome scanning (Leppek et al., 2022).
- iii. Coding Sequence (CDS): The open reading frame that specifies the amino acid sequence of the protein. Codon usage within the CDS can affect translation speed and fidelity (Gustafsson, Govindarajan, & Minshull, 2004).
- iv. 3' Untranslated Region (UTR): A critical hub for post-transcriptional regulation, containing binding sites for microRNAs (miRNAs) and RNA-binding proteins (RBPs) that govern mRNA stability, localization, and translation. AU-rich elements (AREs) in 3' UTRs are classic destabilizing motifs.
- v. Poly(A) Tail: A stretch of adenosines at the 3' end, added by poly(A) polymerase. It protects against 3' exonuclease degradation and synergizes with the 5' cap to enhance translation by promoting circularization of the mRNA via the cap-binding complex (eIF4F) and poly(A)-binding protein (PABP).

2. mRNA Biogenesis: From Transcription to Maturation

mRNA production is a tightly coordinated, multi-step process:

- i. Transcription: RNA Polymerase II synthesizes a precursor mRNA (pre-mRNA).
- ii. 5' Capping: The 5' cap is added co-transcriptionally.
- iii. Splicing: The spliceosome removes non-coding introns and ligates exons. Alternative splicing generates multiple protein isoforms from a single gene.
- iv. 3' End Processing and Polyadenylation: The pre-mRNA is cleaved, and the poly(A) tail is added.
- v. Nuclear Export: The mature mRNA, bound by export factors, is transported through nuclear pore complexes to the cytoplasm.
- vi. Quality Control: Surveillance mechanisms like nonsense-mediated decay (NMD) detect and destroy mRNAs with premature stop codons, preventing the production of truncated proteins.

3. Regulation of mRNA Fate and Translation

mRNA levels and translation are dynamically controlled. Cytoplasmic mRNA half-lives range from minutes to hours. Key regulators include:

- i. miRNAs: Short non-coding RNAs that bind to complementary sequences in the 3' UTR, typically leading to translational repression and mRNA deadenylation/decay.
- ii. RNA-Binding Proteins (RBPs): Hundreds of RBPs bind to specific motifs in UTRs, forming ribonucleoprotein complexes that dictate the mRNA's fate—its stability, localization to specific subcellular compartments (e.g., axons, dendrites), and translation rate in response to cellular signals.

Engineering Synthetic mRNA: From IVT to a Refined Drug Substance

1. *In Vitro* Transcription (IVT): The Production Engine

Synthetic mRNA is produced enzymatically in a cell-free system, a process standardized from molecular biology techniques (Beckert & Masquida, 2011). The reaction requires a linearized DNA template containing a bacteriophage promoter (T7, SP6, or T3) followed by the desired sequence: optimized 5' UTR, codon-optimized CDS, 3' UTR, and a poly(dT) tract for *in vitro* polyadenylation (Pardi et al., 2018). The core components are bacteriophage RNA polymerase, nucleotide triphosphates (NTPs), and a capping strategy. Early methods used cap analogs like the Anti-Reverse Cap Analog (ARCA) added co-transcriptionally to ensure proper orientation and prevent reverse incorporation (Stepinski, Waddell, Stolarski, Darzynkiewicz, & Rhoads, 2001). However, the industry standard has shifted toward post-transcriptional enzymatic capping using vaccinia virus capping enzyme and 2'-O-methyltransferase to generate the Cap 1 structure (7mGpppN1m-), which is naturally recognized by eukaryotic translation initiation factor 4E (eIF4E) and is significantly less immunogenic than Cap 0 structures (Henderson et al., 2021).

2. Key Modifications for Therapeutic Efficacy

The innate immune system is exquisitely tuned to detect viral RNA through pattern recognition receptors (PRRs) like Toll-like receptors (TLR3, TLR7, TLR8) and cytosolic sensors (RIG-I, MDA5). Unmodified IVT mRNA is a potent ligand for these receptors, leading to interferon (IFN) activation and a global shutdown of translation—the very process needed for therapeutic efficacy (Alexopoulou, Holt, Medzhitov, & Flavell, 2001; Hornung et al., 2006). The field's pivotal breakthrough was the demonstration by Karikó and Weissman that incorporating naturally occurring modified nucleosides, specifically pseudouridine (Ψ) or N1-methylpseudouridine (m1 Ψ), into IVT mRNA dramatically reduced activation of TLRs and protein kinase R (PKR) (Karikó et al., 2005, 2008). This suppression of the innate immune response led to a substantial increase in protein expression in mammalian cells by preventing translational inhibition and mRNA degradation (Anderson et al., 2011). Beyond immunomodulation, nucleoside modifications can also enhance translational fidelity and stability (Eyler et al., 2019).

Sequence Optimization is equally critical. Codon optimization, which replaces rare codons with synonymous, host-preferred codons, enhances translational efficiency by matching the abundant tRNA pool, thereby increasing protein yield without altering the amino acid sequence (Gustafsson et al., 2004). UTR engineering involves replacing native UTRs with well-characterized, stable UTRs from highly expressed genes (e.g., human α -globin and β -globin) to provide predictable, high-level translation (Asrani et al., 2018). Furthermore, optimizing GC content and minimizing complex secondary structures in the 5' UTR can facilitate more efficient ribosome scanning and initiation (Leppek et al., 2022).

Purification is a final, critical step to remove immunogenic byproducts of the IVT reaction, particularly double-stranded RNA (dsRNA) contaminants, which are potent activators of MDA5 and PKR (Weissman, Pardi, Muramatsu, & Karikó, 2013). High-performance liquid chromatography (HPLC) and cellulose-based purification methods have become standard for producing clinical-grade mRNA with minimal dsRNA content (Baiersdörfer et al., 2019).

3. Advanced mRNA Formats

- i. Self-Amplifying mRNA (saRNA): Derived from the genome of positive-sense RNA viruses like alphaviruses, saRNA encodes both the antigen of interest and a viral replicase complex (e.g., nsP1-4). Upon delivery, the replicase amplifies the RNA strand intracellularly, leading to much

- higher and more prolonged antigen expression from a dramatically lower initial dose compared to conventional mRNA (Geall et al., 2012). However, this comes with increased complexity, a larger payload size (~9-12 kb), and inherent immunogenicity from the replicase itself (Bloom, van den Berg, & Arbutnot, 2021).
- ii. Circular RNA (circRNA): Engineered as covalently closed, single-stranded loops without free 5' or 3' ends, circRNAs are resistant to exonuclease-mediated decay (Chen & Wang, 2022). This architecture offers the potential for extremely long-lasting protein expression (weeks to months) from a single administration. A major challenge has been enabling cap-independent translation, often solved by incorporating internal ribosome entry site (IRES) elements or engineering N6-methyladenosine (m6A) sites to recruit initiation factors (Wesselhoeft et al., 2019).

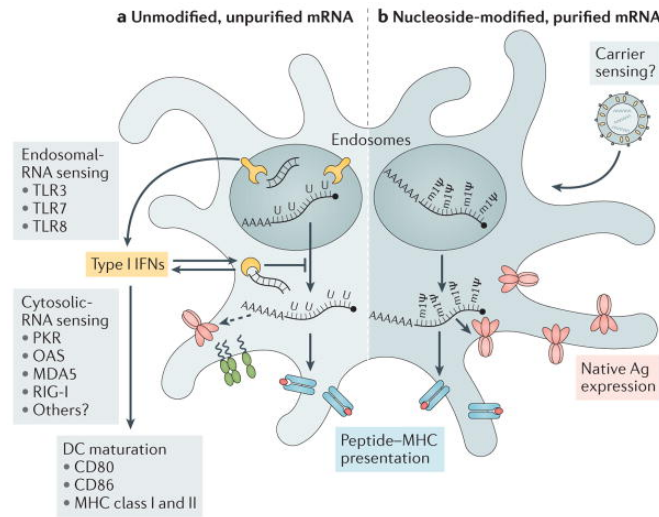


Figure 1. Innate immune sensing of mRNA vaccines

Innate immune sensing of two types of mRNA vaccine by a dendritic cell (DC), with RNA sensors shown in yellow, antigen in red, DC maturation factors in green, and peptide-major histocompatibility complex (MHC) complexes in light blue and red; an example lipid nanoparticle carrier is shown at the top right. A non-exhaustive list of the major known RNA sensors that contribute to the recognition of double-stranded and unmodified single-stranded RNAs is shown. Unmodified, unpurified (part a) and nucleoside-modified, fast protein liquid chromatography (FPLC)-purified (part b) mRNAs were selected for illustration of two formats of mRNA vaccines where known forms of mRNA sensing are present and absent, respectively. The dashed arrow represents reduced antigen expression. Ag, antigen; PKR, interferon-induced, double-stranded RNA-activated protein kinase; MDA5, interferon-induced helicase C domain-containing protein 1 (also known as IFIH1); IFN, interferon; m1Ψ, 1-methylpseudouridine; OAS, 2'-5'-oligoadenylate synthetase; TLR, Toll-like receptor. Figure (1) Engineering and delivery of synthetic mRNA. Schematic created using BioRender.com, incorporating design concepts from Pardi et al. (2018) and delivery mechanisms from Hou et al. (2021).

Delivery Technologies: The Bridge to Clinical Reality

1. The Delivery Imperative

Naked mRNA is rapidly degraded by extracellular ribonucleases (RNases), cannot cross the anionic phospholipid bilayer of cell membranes due to its large size and negative charge, and is sequestered in endosomes after endocytosis, destined for lysosomal degradation (Dowdy, 2017). An effective delivery system must therefore fulfill three key functions: (1) protect the mRNA cargo during systemic transit, (2) facilitate cellular uptake, and (3) enable endosomal escape to release the functional mRNA into the cytosol for translation (Hou et al., 2021).

2. Lipid Nanoparticles (LNPs): The Leading Platform

The clinical success of mRNA vaccines and therapies is inextricably linked to the development of safe and effective LNPs. Modern LNPs are sophisticated, multi-component systems (Cullis & Hope, 2017):

- i. Ionizable Lipid: The most critical functional component. It is cationic at low pH (aiding mRNA encapsulation) and neutrally charged at physiological pH (reducing toxicity). In the acidic

Copyright: Copyrights retained to the Authors. Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

environment of the endosome, it becomes protonated, enabling interaction with anionic endosomal lipids to induce membrane destabilization and pore formation, facilitating mRNA release (Semple et al., 2010). Key examples include DLin-MC3-DMA (used in the first approved siRNA drug, Onpatro), SM-102 (Moderna's COVID-19 vaccine), and ALC-0315 (Pfizer-BioNTech's COVID-19 vaccine) (Corbett et al., 2020; Hassett et al., 2021).

- ii. Phospholipid (e.g., Distearoylphosphatidylcholine, DSPC): Provides structural integrity to the LNP bilayer, contributing to stability and fusion characteristics.
- iii. Cholesterol: Stabilizes the LNP bilayer structure and enhances membrane fluidity and fusion capacity.
- iv. PEGylated Lipid: A polyethylene glycol (PEG)-conjugated lipid that shields the particle surface, modulates particle size, prevents aggregation, and reduces nonspecific protein adsorption and rapid clearance by the mononuclear phagocyte system (MPS). A significant drawback is the potential induction of anti-PEG antibodies, which can cause accelerated blood clearance and reduced efficacy upon repeated dosing (Abu Lila, Kiwada, & Ishida, 2013).

LNPs are typically formulated via rapid mixing of an ethanol phase containing lipids with an aqueous phase containing mRNA in a microfluidic device, producing particles of ~80-100 nm with high encapsulation efficiency (>90%) (Belliveau et al., 2012).

3. Targeting and Route of Administration

Following intravenous administration, current LNPs predominantly accumulate in the liver due to apolipoprotein E (ApoE) adsorption and subsequent uptake by hepatocytes via low-density lipoprotein receptor (LDLR) mediated endocytosis (Akinc et al., 2019). For applications beyond hepatocytes, active targeting strategies are under intense investigation. This includes engineering LNPs with different lipid chemistries to alter organ tropism (e.g., to lung or spleen), or decorating their surface with targeting ligands such as antibodies, peptides, or small molecules to direct them to specific cell types (e.g., immune cells, endothelial cells) (Cheng et al., 2020). The route of administration itself is a powerful targeting tool; intramuscular injection localizes expression primarily to muscle and resident antigen-presenting cells, while intratumoral or intracranial injection directly targets the disease site.

4. Alternative Delivery Systems

While LNPs dominate, other platforms are being explored:

- i. Polymeric Nanoparticles: Using cationic or ionizable polymers like polyethylenimine (PEI) or biodegradable poly(beta-amino esters) (PBAEs) that complex mRNA via electrostatic interactions (Kowalski, Rudra, Miao, & Anderson, 2019).
- ii. Peptide-Based Systems: Cell-penetrating peptides (CPPs) or fusogenic peptides designed to condense mRNA and enhance cellular uptake and endosomal escape (Udhayakumar et al., 2021).
- iii. Conjugate Technologies: Direct covalent conjugation of mRNA to targeting ligands (e.g., GalNAc for hepatocyte targeting) or polymers to improve stability and pharmacokinetics (Springer & Dowdy, 2018).

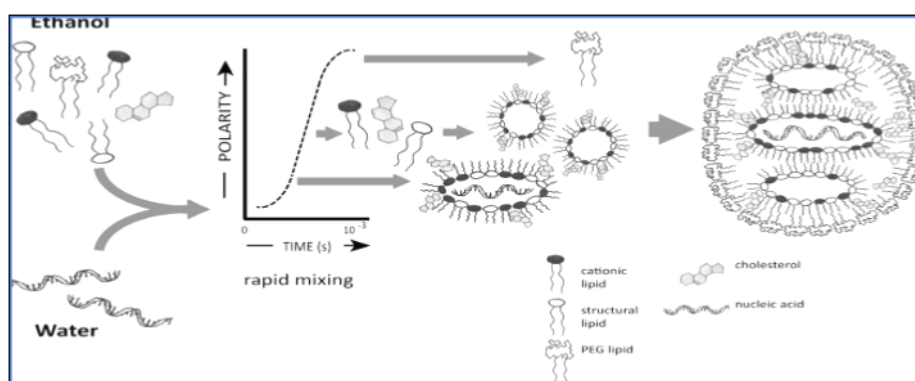


Fig 2: Ethanol Loading Formulation Process for LNP Containing Oligonucleotides Such as siRNA. Figure (3) Engineering and delivery of synthetic mRNA. Schematic created using BioRender.com, incorporating design concepts from Cullis & Hope, (2017).

Therapeutic Applications: An Expanding Universe

1. Prophylactic Vaccines

- i. Infectious Diseases: Beyond COVID-19, active clinical programs for influenza (seeking improved breadth and durability), RSV, HIV, Zika, Nipah, and Epstein-Barr Virus. Advantages: rapid response to pandemic threats and variant updates.
- ii. Generalizable Advantage: mRNA vaccines induce strong CD4⁺ T cell, CD8⁺ T cell, and neutralizing antibody responses. LNPs have intrinsic adjuvant properties, potentially activating follicular helper T cells and germinal center B cell responses (Laczko et al., 2020).

2. Therapeutic Vaccines

- i. Oncology (Cancer Immunotherapy): Personalized Neoantigen Vaccines: Tumor sequencing identifies patient-specific mutations. mRNA encoding these neoantigens is manufactured and administered to prime T cells to attack the tumor. Promising data in melanoma (Moderna/Merck) (Sahin et al., 2020). Also, vaccines for shared tumor-associated antigens (e.g., TAA, CEA).
- ii. Other: Therapeutic vaccines for chronic infections (e.g., herpes simplex virus, hepatitis B).

3. Protein Replacement and Regenerative Therapy

- i. *In vivo* Protein Production: mRNA acts as a temporary blueprint to produce proteins inside the patient's own cells, overcoming challenges of recombinant protein manufacturing, stability, and delivery.
- ii. Rare Diseases: Clinical trials for methylmalonic acidemia (propionic enzyme), cystic fibrosis (CFTR protein), glycogen storage disease (Rohner et al., 2022).
- iii. Regenerative Medicine: mRNA encoding VEGF for angiogenesis in heart disease; BMP-2 for bone growth; factors for tissue repair (Zangi et al., 2013).
- iv. Advantage: Transient expression is ideal for many signaling proteins, reducing risks of genomic integration or long-term overexpression.

4. Gene Editing and Cellular Reprogramming

- i. Non-viral CRISPR-Cas9 Delivery: mRNA encoding the Cas9 nuclease (and a separate guide RNA) allows transient, high-efficiency expression of the editing machinery, significantly reducing off-target risks compared to stable viral expression. Used *ex vivo* (engineer CAR-T cells) and *in vivo* (e.g., for transthyretin amyloidosis) (Finn et al., 2018).
- ii. Cell Fate Reprogramming: mRNA cocktails of transcription factors can directly reprogram somatic cells (e.g., fibroblasts into cardiomyocytes or neurons) for regenerative purposes (Warren et al., 2010).

Challenges and Future Perspectives

1. Persistent Challenges

- i. Precision Delivery: Achieving efficient, specific delivery to non-liver tissues (e.g., lungs, heart, brain, specific immune cells) remains a primary hurdle (Dammes & Peer, 2020).
- ii. Durability & Redosing: For many chronic conditions, protein expression from current mRNA lasts days to a week. Solutions include saRNA, circRNA, or improved formulations. Anti-PEG immunity and anti-drug antibodies can limit repeat dosing.
- iii. Scalability & Cost: While scalable, GMP manufacturing of mRNA-LNPs is complex. Reducing cost is critical for global health equity (Kis, Shah, & Sato, 2022).
- iv. Long-Term Safety: Continued pharmacovigilance is essential. Areas of monitoring include: rare adverse events (e.g., myocarditis), lipid carrier toxicology, and long-term immunological effects of repeated LNP administration.

2. Future Directions

- i. Next-Generation Constructs: Clinical translation of circRNA and optimized saRNA for durable expression.

- ii. Programmable Therapeutics: "Smart" mRNA systems responsive to cellular cues or small molecules for controlled protein expression.

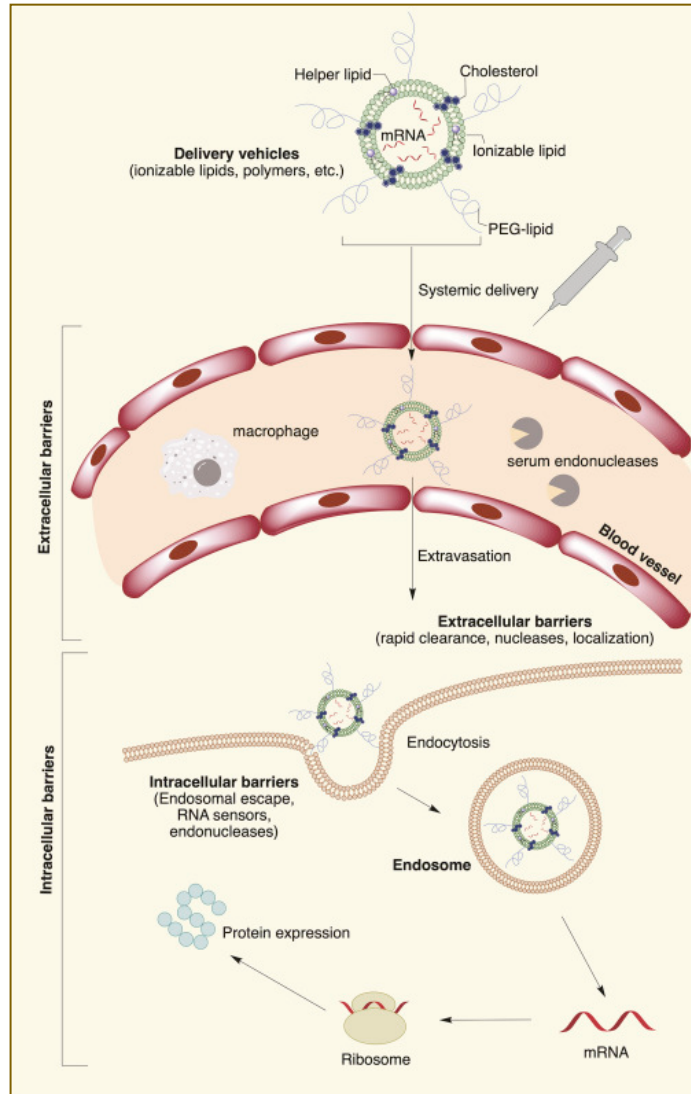


Figure 3 Schematic Representation of Extra- and Intracellular Barriers for mRNA Delivery Figure (3) Engineering and delivery of synthetic mRNA. Schematic created using BioRender.com, incorporating design concepts from Kowalski et al., (2019).

- iii. Disease Prevention: Potential for multi-valent pandemic-preparedness vaccines or routine cancer prevention vaccines (e.g., for KRAS-mutant pre-cancers).
- iv. Integration with Other Modalities: Combining mRNA vaccines with checkpoint inhibitors in oncology, or mRNA-encoded antibodies with small molecules.
- v. Expansion into New Diseases: Neurological disorders, autoimmune diseases, metabolic conditions, and more.

Conclusion

The mRNA technology platform has irrevocably changed the landscape of medicine. Its journey from a fundamental biological concept to a validated clinical powerhouse is a testament to decades of basic science and persistent innovation. By harnessing and refining the cell's own translational machinery, mRNA therapeutics offer a unique combination of speed, flexibility, efficacy, and manufacturability. While challenges in delivery, durability, and cost remain active frontiers of research, the trajectory is clear. mRNA is not a one-pandemic

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net

wonder but a foundational pillar of 21st-century biomedicine, poised to deliver a new generation of treatments for some of humanity's most intractable diseases. The future will be written, in part, in the language of messenger RNA.

Conflicts of Interest

The authors declare no conflicts of interest.

Acknowledgment

We would like to thank the National Research Centre, Egypt.

References

- Abu Lila, A. S., Kiwada, H., & Ishida, T. (2013). The accelerated blood clearance (ABC) phenomenon: Clinical challenge and approaches to manage. *Journal of Controlled Release*, 172(1), 38–47. <https://doi.org/10.1016/j.jconrel.2013.07.026>
- Akinc, A., Maier, M. A., Manoharan, M., Fitzgerald, K., Jayaraman, M., Barros, S., Ansell, S., Du, X., Hope, M. J., Madden, T. D., Mui, B. L., Semple, S. C., Tam, Y. K., Ciufolini, M., Witzigmann, D., Kulkarni, J. A., van der Meel, R., & Cullis, P. R. (2019). The Onpattro story and the clinical translation of nanomedicines containing nucleic acid-based drugs. *Nature Nanotechnology*, 14(12), 1084–1087. <https://doi.org/10.1038/s41565-019-0591-y>
- Alexopoulou, L., Holt, A. C., Medzhitov, R., & Flavell, R. A. (2001). Recognition of double-stranded RNA and activation of NF- κ B by Toll-like receptor 3. *Nature*, 413(6857), 732–738. <https://doi.org/10.1038/35099560>
- Anderson, B. R., Muramatsu, H., Nallagatla, S. R., Bevilacqua, P. C., Sansing, L. H., Weissman, D., & Karikó, K. (2011). Incorporation of pseudouridine into mRNA enhances translation by diminishing PKR activation. *Nucleic Acids Research*, 38(17), 5884–5892. <https://doi.org/10.1093/nar/gkr347>
- Asrani, K. H., Farelli, J. D., Stahley, M. R., Miller, R. L., Cheng, C. J., Subramanian, R. R., & Brown, J. M. (2018). Optimization of mRNA untranslated regions for improved expression of therapeutic mRNA. *RNA Biology*, 15(6), 756–762. <https://doi.org/10.1080/15476286.2018.1450054>
- Baiersdörfer, M., Boros, G., Muramatsu, H., Mahiny, A., Vlatkovic, I., Sahin, U., & Karikó, K. (2019). A facile method for the removal of dsRNA contaminant from *in vitro*-transcribed mRNA. *Molecular Therapy - Nucleic Acids*, 15, 26–35. <https://doi.org/10.1016/j.omtn.2019.02.018>
- Beckert, B., & Masquida, B. (2011). Synthesis of RNA by *in vitro* transcription. In *RNA* (pp. 29–41). Humana Press. https://doi.org/10.1007/978-1-59745-248-9_3
- Belliveau, N. M., Huft, J., Lin, P. J., Chen, S., Leung, A. K., Leaver, T. J., Wild, A. W., Lee, J. B., Taylor, R. J., Tam, Y. K., Hansen, C. L., & Cullis, P. R. (2012). Microfluidic synthesis of highly potent limit-size lipid nanoparticles for *in vivo* delivery of siRNA. *Molecular Therapy - Nucleic Acids*, 1, e37. <https://doi.org/10.1038/mtna.2012.28>
- Bloom, K., van den Berg, F., & Arbuthnot, P. (2021). Self-amplifying RNA vaccines for infectious diseases. *Gene Therapy*, 28(3–4), 117–129. <https://doi.org/10.1038/s41434-020-00204-y>
- Brenner, S., Jacob, F., & Meselson, M. (1961). An unstable intermediate carrying information from genes to ribosomes for protein synthesis. *Nature*, 190, 576–581. <https://doi.org/10.1038/190576a0>
- Chen, R., & Wang, S. K. (2022). Circular RNAs in physiology and non-immunological diseases. *Trends in Biochemical Sciences*, 47(3), 250–264. <https://doi.org/10.1016/j.tibs.2021.08.006>

Cheng, Q., Wei, T., Jia, Y., Farbiak, L., Zhou, K., Zhang, S., Wei, Y., Zhu, H., & Siegwart, D. J. (2020). Dendrimer-based lipid nanoparticles deliver therapeutic FAH mRNA to normalize liver function and extend survival in a mouse model of hepatorenal tyrosinemia type I. *Advanced Materials*, 32(52), 2003537. <https://doi.org/10.1002/adma.202003537>

Corbett, K. S., Edwards, D. K., Leist, S. R., Abiona, O. M., Boyoglu-Barnum, S., Gillespie, R. A., Himansu, S., Schäfer, A., Ziwawo, C. T., DiPiazza, A. T., Dinnon, K. H., Elbashir, S. M., Shaw, C. A., Woods, A., Fritch, E. J., Martinez, D. R., Bock, K. W., Minai, M., Nagata, B. M., ... Graham, B. S. (2020). SARS-CoV-2 mRNA vaccine design enabled by prototype pathogen preparedness. *Nature*, 586(7830), 567–571. <https://doi.org/10.1038/s41586-020-2622-0>

Cullis, P. R., & Hope, M. J. (2017). Lipid nanoparticle systems for enabling gene therapies. *Molecular Therapy*, 25(7), 1467–1475. <https://doi.org/10.1016/j.ymthe.2017.03.013>

Dammes, N., & Peer, D. (2020). Paving the road for RNA therapeutics. *Trends in Pharmacological Sciences*, 41(10), 755–775. <https://doi.org/10.1016/j.tips.2020.08.004>

Dowdy, S. F. (2017). Overcoming cellular barriers for RNA therapeutics. *Nature Biotechnology*, 35(3), 222–229. <https://doi.org/10.1038/nbt.3802>

Eyler, D. E., Franco, M. K., Batool, Z., Wu, M. Z., Dubuke, M. L., Dobosz-Bartoszek, M., Jones, J. D., Polikanov, Y. S., Roy, B., & Koutmou, K. S. (2019). Pseudouridylation of mRNA coding sequences alters translation. *Proceedings of the National Academy of Sciences*, 116(46), 23068–23074. <https://doi.org/10.1073/pnas.1821754116>

Finn, J. D., Smith, A. R., Patel, M. C., Shaw, L., Youniss, M. R., van Heteren, J., Dirstine, T., Ciullo, C., Lescarbeau, R., Seitzer, J., Shah, R. R., Shah, A., Ling, D., Growe, J., Pink, M., Rohde, E., Wood, K. M., Salomon, W. E., Harrington, W. F., ... Morrissey, D. V. (2018). A single administration of CRISPR/Cas9 lipid nanoparticles achieves robust and persistent *in vivo* genome editing. *Cell Reports*, 22(9), 2227–2235. <https://doi.org/10.1016/j.celrep.2018.02.014>

Geall, A. J., Verma, A., Otten, G. R., Shaw, C. A., Hekele, A., Banerjee, K., Cu, Y., Beard, C. W., Brito, L. A., Krucker, T., O'Hagan, D. T., Singh, M., Mason, P. W., Valiante, N. M., Dormitzer, P. R., Barnett, S. W., Rappuoli, R., Ulmer, J. B., & Mandl, C. W. (2012). Nonviral delivery of self-amplifying RNA vaccines. *Proceedings of the National Academy of Sciences*, 109(36), 14604–14609. <https://doi.org/10.1073/pnas.1209367109>

Gustafsson, C., Govindarajan, S., & Minshull, J. (2004). Codon bias and heterologous protein expression. *Trends in Biotechnology*, 22(7), 346–353. <https://doi.org/10.1016/j.tibtech.2004.04.006>

Hassett, K. J., Benenato, K. E., Jacquinet, E., Lee, A., Woods, A., Yuzhakov, O., Himansu, S., Deterling, J., Geilich, B. M., Ketova, T., Mihai, C., Lynn, A., McFadyen, I., Moore, M. J., Senn, J. J., Stanton, M. G., Almarsson, Ö., Ciaramella, G., & Brito, L. A. (2021). Optimization of lipid nanoparticles for intramuscular administration of mRNA vaccines. *Molecular Therapy - Nucleic Acids*, 25, 1–11. <https://doi.org/10.1016/j.omtn.2021.07.003>

Henderson, J. M., Ujita, A., Hill, E., Yousif-Rosales, S., Smith, C., Ko, N., McReynolds, T., Cabral, C. R., Escamilla-Powers, J. R., & Houston, M. E. (2021). Cap 1 messenger RNA synthesis with co-transcriptional CleanCap® analog by *in vitro* transcription. *Current Protocols*, 1(2), e39. <https://doi.org/10.1002/cpz1.39>

Hornung, V., Ellegast, J., Kim, S., Brzózka, K., Jung, A., Kato, H., Poeck, H., Akira, S., Conzelmann, K. K., Schlee, M., Endres, S., & Hartmann, G. (2006). 5'-Triphosphate RNA is the ligand for RIG-I. *Science*, 314(5801), 994–997. <https://doi.org/10.1126/science.1132505>

Copyright: Copyrights retained to the Authors. Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

Hou, X., Zaks, T., Langer, R., & Dong, Y. (2021). Lipid nanoparticles for mRNA delivery. *Nature Reviews Materials*, 6(12), 1078–1094. <https://doi.org/10.1038/s41578-021-00358-0>

Karikó, K., Buckstein, M., Ni, H., & Weissman, D. (2005). Suppression of RNA recognition by Toll-like receptors: The impact of nucleoside modification and the evolutionary origin of RNA. *Immunity*, 23(2), 165–175. <https://doi.org/10.1016/j.immuni.2005.06.008>

Karikó, K., Muramatsu, H., Welsh, F. A., Ludwig, J., Kato, H., Akira, S., & Weissman, D. (2008). Incorporation of pseudouridine into mRNA yields superior nonimmunogenic vector with increased translational capacity and biological stability. *Molecular Therapy*, 16(11), 1833–1840. <https://doi.org/10.1038/mt.2008.200>

Kis, Z., Shah, N., & Sato, Y. (2022). The future of vaccine manufacturing: Leveraging disruptive technologies to improve global health. *Journal of Pharmaceutical Sciences*, 111(5), 1241–1249. <https://doi.org/10.1016/j.xphs.2021.12.013>

Kowalski, P. S., Rudra, A., Miao, L., & Anderson, D. G. (2019). Delivering the messenger: Advances in technologies for therapeutic mRNA delivery. *Molecular Therapy*, 27(4), 710–728. <https://doi.org/10.1016/j.ymthe.2019.02.012>

Laczko, D., Hogan, M. J., Toulmin, S. A., Hicks, P., Lederer, K., Gaudette, B. T., Castaño, D., Amanat, F., Muramatsu, H., Oguin, T. H., Ojha, A., Zhang, L., Mu, Z., Parks, R., Manzoni, T. B., Roper, B., Strohmeier, S., Tombác, I., Arwood, L., ... Weissman, D. (2020). A single immunization with nucleoside-modified mRNA vaccines elicits strong cellular and humoral immune responses against SARS-CoV-2 in mice. *Immunity*, 53(4), 724-732.e7. <https://doi.org/10.1016/j.immuni.2020.07.019>

Leppek, K., Byeon, G. W., Kladwang, W., Wayment-Steele, H. K., Kerr, C. H., Xu, A. F., Kim, D. S., Topkar, V. V., Choe, C., Rothschild, D., Tiu, G. C., Wellington-Oguri, R., Fujii, K., Sharma, E., Watkins, A. M., Nicol, J. J., Romano, J., Tunguz, B., Diaz, F., ... Das, R. (2022). Combinatorial optimization of mRNA structure, stability, and translation for RNA-based therapeutics. *Nature Communications*, 13(1), 1536. <https://doi.org/10.1038/s41467-022-28776-w>

Pardi, N., Hogan, M. J., Porter, F. W., & Weissman, D. (2018). mRNA vaccines — a new era in vaccinology. *Nature Reviews Drug Discovery*, 17(4), 261–279. <https://doi.org/10.1038/nrd.2017.243>

Polack, F. P., Thomas, S. J., Kitchin, N., Absalon, J., Gurtman, A., Lockhart, S., Perez, J. L., Pérez Marc, G., Moreira, E. D., Zerbini, C., Bailey, R., Swanson, K. A., Roychoudhury, S., Koury, K., Li, P., Kalina, W. V., Cooper, D., Frenck, R. W., Hammitt, L. L., ... Gruber, W. C. (2020). Safety and Efficacy of the BNT162b2 mRNA Covid-19 Vaccine. *New England Journal of Medicine*, 383(27), 2603–2615. <https://doi.org/10.1056/NEJMoa2034577>

Rohner, E., Yang, R., Foo, K. S., Goedel, A., & Chien, K. R. (2022). Unlocking the promise of mRNA therapeutics. *Nature Biotechnology*, 40(11), 1586–1600. <https://doi.org/10.1038/s41587-022-01491-z>

Sahin, U., Oehm, P., Derhovanessian, E., Jabulowsky, R. A., Vormehr, M., Gold, M., Maurus, D., Schwarck-Kokarakis, D., Kuhn, A. N., Omokoko, T., Kranz, L. M., Diken, M., Kreiter, S., Haas, H., Attig, S., Rae, R., Cuk, K., Kemmer-Brück, A., Breikreuz, A., ... Türeci, Ö. (2020). An RNA vaccine drives immunity in checkpoint-inhibitor-treated melanoma. *Nature*, 585(7823), 107–112. <https://doi.org/10.1038/s41586-020-2537-8>

Semple, S. C., Akinc, A., Chen, J., Sandhu, A. P., Mui, B. L., Cho, C. K., Sah, D. W., Stebbing, D., Crosley, E. J., Yaworski, E., Hafez, I. M., Dorkin, J. R., Qin, J., Lam, K., Rajeev, K. G., Wong, K. F., Jeffs, L. B., Nechev, L., Eisenhardt, M. L., ... Hope, M. J. (2010). Rational design of cationic lipids for siRNA delivery. *Nature Biotechnology*, 28(2), 172–176. <https://doi.org/10.1038/nbt.1602>

Copyright: Copyrights retained to the Authors. Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

Springer, A. D., & Dowdy, S. F. (2018). GalNAc-siRNA conjugates: Leading the way for delivery of RNAi therapeutics. *Nucleic Acid Therapeutics*, 28(3), 109–118. <https://doi.org/10.1089/nat.2018.0736>

Stepinski, J., Waddell, C., Stolarski, R., Darzynkiewicz, E., & Rhoads, R. E. (2001). Synthesis and properties of mRNAs containing the novel "anti-reverse" cap analogs 7-methyl(3'-O-methyl)GpppG and 7-methyl (3'-deoxy)GpppG. *RNA*, 7(10), 1486–1495. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1370193/>

Udhayakumar, V. K., De Beuckelaer, A., McCaffrey, J., McCrudden, C. M., Kirschman, J. L., Vanover, D., Van Hoecke, L., Roose, K., Deswarte, K., De Geest, B. G., Lienenklaus, S., & De Koker, S. (2021). Arginine-rich peptide-based mRNA nanocomplexes efficiently instigate cytotoxic T cell immunity dependent on the amphipathic organization of the peptide. *Advanced Healthcare Materials*, 10(7), 2001652. <https://doi.org/10.1002/adhm.202001652>

Warren, L., Manos, P. D., Ahfeldt, T., Loh, Y.-H., Li, H., Lau, F., Ebina, W., Mandal, P. K., Smith, Z. D., Meissner, A., Daley, G. Q., Brack, A. S., Collins, J. J., Cowan, C., Schlaeger, T. M., & Rossi, D. J. (2010). Highly efficient reprogramming to pluripotency and directed differentiation of human cells with synthetic modified mRNA. *Cell Stem Cell*, 7(5), 618–630. <https://doi.org/10.1016/j.stem.2010.08.012>

Weissman, D., Pardi, N., Muramatsu, H., & Karikó, K. (2013). HPLC purification of *in vitro* transcribed long RNA. *Methods in Molecular Biology*, 969, 43–54. https://doi.org/10.1007/978-1-62703-260-5_3

Wesselhoeft, R. A., Kowalski, P. S., Parker-Hale, F. C., Huang, Y., Biswas, S., & Anderson, D. G. (2019). RNA circularization diminishes immunogenicity and can extend translation duration *in vivo*. *Molecular Cell*, 74(3), 508-520.e4. <https://doi.org/10.1016/j.molcel.2019.02.015>

Wolff, J. A., Malone, R. W., Williams, P., Chong, W., Acsadi, G., Jani, A., & Felgner, P. L. (1990). Direct gene transfer into mouse muscle *in vivo*. *Science*, 247(4949 Pt 1), 1465–1468. <https://doi.org/10.1126/science.1690918>

Zangi, L., Lui, K. O., von Gise, A., Ma, Q., Ebina, W., Ptaszek, L. M., Später, D., Xu, H., Tabebordbar, M., Gorbатов, R., Sena, B., Nahrendorf, M., Briscoe, D. M., Li, R. A., Wagers, A. J., Rossi, D. J., Pu, W. T., & Chien, K. R. (2013). Modified mRNA directs the fate of heart progenitor cells and induces vascular regeneration after myocardial infarction. *Nature Biotechnology*, 31(10), 898–907. <https://doi.org/10.1038/nbt.2682>

The Epidemiological Nexus: Age, Geography, and Infectious Disease in Egyptian Cattle

Age and Geography of Cattle Diseases in Egypt

Abouelhag H. A. *

*Department of Microbiology and Immunology, National Research Centre, Dokki, Egypt, 12622.

Received: 19-12-2025

Accepted: 10-01-2026

Published online: 30-01-2026

DOI: <https://doi.org/10.33687/ricosbiol.04.01.105>

Abstract

Infectious diseases impose a severe constraint on cattle productivity, food security, and the national economy in Egypt. The susceptibility to, and manifestation of, these diseases are profoundly influenced by the age of the host animal, a factor often overlooked in broad control strategies. This review systematically synthesizes the scientific literature on Egypt's major bovine infectious diseases to elucidate the critical correlation between host age and disease epidemiology and to map the associated geographical distribution patterns. Our analysis reveals distinct age-specific syndromes: neonates (0-1 month) are dominated by enteric pathogens like *E. coli* and *Cryptosporidium*; growing calves (1-12 months) are most susceptible to severe outcomes from foot-and-mouth disease (FMD) and bovine viral diarrhoea virus (BVDV) persistence; while adult cattle are most impacted by production diseases such as brucellosis, mastitis, and chronic fascioliasis. Geographically, the Nile Delta is identified as a multifactorial hotspot due to high animal density, intensive farming, and irrigation networks, whereas Upper Egypt and newly reclaimed areas exhibit distinct outbreak dynamics. The movement of specific age cohorts, such as pregnant heifers and weaned calves, is a key driver of disease spread. This synthesis underscores the imperative for a dual-targeting control strategy: implementing age-specific interventions within defined geographical risk zones. We conclude with evidence-based recommendations for age-stratified surveillance, vaccination, and management practices tailored to Egypt's diverse farming systems to enhance disease control and livestock productivity.

Keywords: Cattle diseases, Egypt, age susceptibility, geographical distribution, epidemiology, FMD, brucellosis, BVD.

Introduction

Cattle are a cornerstone of Egypt's agricultural economy, vital for dairy, meat, and draft power. However, the livestock sector faces persistent challenges from a range of infectious diseases that cause significant morbidity, mortality, and economic loss (Gamil et al., 2024). The epidemiological landscape of these diseases is complex, shaped by factors such as climate, husbandry practices, animal movement, and host immunity. Among these, the age of the host animal is a fundamental but often underutilized determinant of disease outcome.

Age influences disease susceptibility through immunological maturity, management-related exposure risks (e.g., colostrum intake, housing), and physiological status. A calf, a growing heifer, and a lactating cow represent vastly different epidemiological units within a herd. In Egypt, where farming systems range from traditional smallholder holdings to intensive commercial dairy operations, understanding the interaction between age, disease, and geography is critical for effective control.

This review aims to systematically compile and analyze the available scientific evidence on major infectious diseases of cattle in Egypt. Its specific objectives are to: 1) Establish clear correlations between host age and the occurrence, clinical presentation, and outcome of key diseases; 2) Map the geographical distribution of these diseases and identify high-risk zones; and 3) Synthesize this information to provide actionable, age- and geography-targeted recommendations for policymakers, veterinarians, and farmers.

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



Methodology

A systematic approach was employed to identify, select, and analyze relevant literature. Electronic databases searched included PubMed, Scopus, Google Scholar, and the Egyptian Knowledge Bank (EKB). The search strategy combined keywords related to cattle ("bovine," "cattle," "Egypt"), specific diseases ("foot-and-mouth disease," "brucellosis," "mastitis," etc.), and epidemiological factors ("age," "susceptibility," "prevalence," "geographical distribution," "Egyptian governorate"). The search was limited to articles published between 2000 and 2024, with a focus on more recent studies where available. Inclusion criteria encompassed original research articles, surveillance reports, and reviews providing data on disease occurrence, age distribution, or geographical location within Egypt. Articles lacking specific age-related data or clear geographical referencing within Egypt were excluded. Data extracted from each study included: disease/pathogen, study location (governorate), sample characteristics (size, age groups), key findings on age association, and reported prevalence.

Selected Major Infectious Diseases

1. Viral Diseases

1.1. Foot-and-Mouth Disease (FMD)

- **Age Correlation:** The clinical and pathological impact of FMD is highly age-dependent. Neonatal and suckling calves (0-3 months) are particularly vulnerable, often suffering high mortality rates (exceeding 70% in outbreaks) due to viral myocarditis, which can occur without the classic signs of vesicular lesions (Soliman et al., 2022). Weaned calves and juveniles (3-12 months) exhibit high morbidity with severe oral and foot lesions, while adult cattle typically show the characteristic vesicles, with economic impact stemming primarily from milk drop and loss of condition.

- **Geographical Distribution & Interface:** FMD is endemic in Egypt with continuous circulation, particularly in the high-density livestock areas of the **Nile Delta** (e.g., Beheira, Gharbia, Dakahlia) (Abdelhakim et al., 2023). Here, the constant exposure pressure means calves are infected at a very young age, leading to endemic neonatal mortality. In contrast, in Upper Egypt governorates (e.g., Minya, Sohag) and newly reclaimed lands (e.g., New Valley, Toshka), outbreaks are often episodic. When the virus enters these areas with more naive populations, explosive outbreaks occur, affecting all age groups severely (Abdelhakim et al., 2023). Serotypes O (EA-3 toptotype), A (African toptotype), and SAT2 have been concurrently circulating, complicating control (Gamil et al., 2024; El-Nahas et al., 2025).

1.2. Bovine Viral Diarrhea Virus (BVDV)

- **Age Correlation:** BVDV epidemiology is intrinsically linked to age via the mechanism of persistent infection (PI). If a pregnant cow is infected between approximately 45-125 days of gestation, the virus can be transmitted to the fetus, resulting in the birth of a PI calf that sheds large amounts of virus for life (Dubovi, 2013). These PI animals are the main reservoir and often succumb to fatal mucosal disease between 6-24 months of age. Acute infection in immunocompetent animals of any age can cause mild enteric or respiratory signs and transient immunosuppression.

- **Geographical Distribution & Interface:** PI animals are disproportionately found in large, intensive dairy herds in the Nile Delta (e.g., Menofia, Qalyubia) due to frequent animal introductions and high population density (Mahmoud et al., 2020). A recent study identified a PI prevalence of 2.5%, with all detected animals being young calves aged 1-6 months, highlighting the age-specific risk window (Ibrahim et al., 2022). The emergence of the HoBi-like pestivirus (BVDV-3) in Egypt further underscores the risk posed by international and regional livestock trade (Ibrahim et al., 2022; El-Damaty et al., 2018).

1.3. Infectious Bovine Rhinotracheitis (IBR) and Lumpy Skin Disease (LSD)

- **IBR** causes severe respiratory disease in weaned calves and feedlot cattle, while in adults it is associated with reproductive failure (abortion, infertility). It is endemic nationwide, with higher seroprevalence in intensive Delta farms.

- **LSD**, which became epidemic in Egypt in 2020, affects all ages. However, high-yielding dairy cows and young calves may experience more severe generalized lesions. Its spread has been nationwide, with cases

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



reported from Alexandria to Aswan, demonstrating rapid geographical dissemination.

2. Bacterial Diseases

2.1. Brucellosis (*Brucella abortus* & *B. melitensis*)

- **Age Correlation:** Clinical disease is tightly coupled to reproductive maturity. The hallmark sign—late-term abortion—is most common in primiparous heifers and adult cows during their first infected pregnancy (Godfroid et al., 2011). Young calves can be infected and may harbor the bacteria for extended periods without showing clinical signs, acting as asymptomatic carriers that disseminate the disease upon reaching breeding age (El-Diasty et al., 2025).

- **Geographical Distribution & Interface:** Brucellosis is endemic, with high seroprevalence reported in both the Nile Delta (Kafr El-Sheikh, Dakahlia) and several Upper Egypt governorates (Assiut, El-Minya) (Khalifa et al., 2018). The risk is amplified in traditional mixed farming systems where cattle, buffalo, and small ruminants share space, facilitating cross-species transmission primarily of *B. melitensis* (El-Diasty et al., 2025). The movement of untested, sub-adult replacement animals from these endemic zones is a primary route of geographic spread to lower-prevalence areas.

2.2. Mastitis and Calf Diarrhea Complex

- **Mastitis:** Primarily a disease of the lactating adult dairy cow, with the peri-parturient period being highest risk. Environmental pathogens (e.g., *E. coli*, *Streptococcus uberis*) can affect all lactating animals, while contagious pathogens like *Staphylococcus aureus* spread within milking herds. It is ubiquitous, with higher prevalence and pathogen diversity in intensive dairy farms of Lower Egypt.

- **Calf Diarrhea:** A neonatal syndrome (0-3 weeks) where age dictates the likely pathogen: enterotoxigenic *E. coli* (K99) in the first week, *Rotavirus*, *Cryptosporidium parvum*, and *Coronavirus* in weeks 1-3, and *Salmonella* spp. potentially later. It is a major cause of pre-weaning mortality across all farming systems, with incidence often linked to failures in colostrum management and hygiene in calf pens.

2.3. Tuberculosis (*Mycobacterium bovis*)

- **Age Correlation:** A chronic, slowly progressive disease. Detection rates are typically higher in older, culled dairy cows due to longer exposure time and the disease's insidious nature. Younger animals may be infected but less frequently show advanced, detectable lesions.

- **Geographical Distribution:** Reported in scattered foci, including Delta governorates (Menofia) and Upper Egypt (Sohag), often linked to intensive farming or areas with historical prevalence. Its zoonotic nature makes it a significant public health concern at the human-animal interface.

3. Parasitic Diseases

3.1. Fascioliasis (*Fasciola gigantica* & *F. hepatica*)

- **Age Correlation:** Infection shows a clear age-prevalence gradient due to cumulative exposure. A 2025 study in the New Valley found prevalence was lowest in animals <1 year old (12.7%) and highest in those >3 years old (30.4%) (Khedr et al., 2025). Young cattle (6-18 months) experiencing first major exposures often develop the most severe chronic, debilitating disease (weight loss, anemia, "bottle jaw").

- **Geographical Distribution & Interface:** The disease is hyperendemic in regions supporting the aquatic snail intermediate host. This includes the canal networks of the Nile Delta and the oasis systems of the New Valley and Fayoum (El-Shahawy & Metwally, 2018; Khedr et al., 2025). The high prevalence in cattle in these areas constitutes a significant zoonotic reservoir for human infection.

3.2. Theileriosis (*Theileria annulata*)

- Affects all ages, but exotic breeds and young calves are most susceptible to severe disease. It is endemic in areas where the *Hyalomma* tick vector is established, primarily the Nile Delta and the New Valley Oases.

4. Synthesis: The Interplay of Age, Geography, and Management

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



Age-Susceptibility Landscape in Egypt:

- **The Neonatal Challenge Belt:** Ubiquitous, driven by enteric pathogens. Mortality is highest in smallholder systems with limited veterinary care.
- **The Growing Stock Risk Zone:** Geographically defined by FMD-endemic areas (Delta) and *Fasciola*-endemic wetlands (Delta, Fayoum, Oases). Economic loss from stunted growth and mortality in this age group is substantial.
- **The Adult Production Loss Zone:** Dominated by mastitis in high-yielding Delta dairy belts and reproductive/chronic diseases (brucellosis, tuberculosis) in older cattle nationwide.

A critical synthesizing factor is animal movement patterns (Fasanmi et al., 2021). The trade of weaned calves spreads BVDV and respiratory pathogens; the movement of pregnant heifers spreads brucellosis; and the importation of livestock introduces new strains of FMDV and BVDV. These movements directly link geographical disease hotspots with susceptible age cohorts in previously lower-risk areas.

Furthermore, farm management systems mediate the age-geography interaction (El Nahas & Hassanain, 2021). Intensive Delta dairies face high mastitis and BVDV challenges in adults and neonates, respectively. Traditional Upper Egyptian systems face higher risks from brucellosis and FMD outbreaks due to different husbandry and trade practices.

Conclusions and Strategic Recommendations

This review demonstrates that a cattle's age is a primary determinant of infectious disease risk in Egypt, and that this risk is spatially heterogeneous. Effective control requires moving beyond blanket strategies to precision interventions.

A. For Policy and National Programs:

1. **Implement Age-Stratified Surveillance:** Disease reporting systems should mandate recording of the age group affected (neonate, juvenile, adult). This data should be integrated into a geospatial platform to visualize and track age-specific disease hotspots in real-time.
2. **Enforce Age- and Risk-Based Movement Controls:** The highest-risk movements (e.g., pregnant heifers, calves from PI-positive herds) should require the most stringent testing and certification. Develop a "test-and-move" protocol based on age and destination risk.
3. **Refine Vaccine Campaigns:** For FMD in endemic Delta regions, prioritize strategies that ensure high maternal immunity to protect neonates (e.g., strategic vaccination of pregnant cows). For brucellosis, continue and strengthen the vaccination of female calves (RB51) before breeding age.

B. For Veterinary Practitioners and Farmers:

1. **Adopt Life-Stage Biosecurity:**
 - **Neonatal Unit:** Ensure colostrum intake within 2 hours, use hygienic, separate calving pens, and implement targeted prophylaxis for calf diarrhea.
 - **Weaner/Grower Unit:** In *Fasciola*-endemic areas, delay first grazing on wet pastures or use strategic pre-grazing anthelmintics. Avoid mixing age groups to reduce respiratory disease spread.
 - **Adult/Lactating Unit:** Focus on mastitis control programs (milking hygiene, dry cow therapy) and regular reproductive health screening.
2. **Implement Targeted "Test-and-Cull":** In BVDV-endemic dairy herds (especially in the Delta), screen all young stock (6-12 months) for PI status and cull positive animals. Screen all breeding-age females for brucellosis before introduction to a clean herd.

C. For Research Priorities:

1. Conduct longitudinal cohort studies in contrasting agro-ecological zones (e.g., Delta village vs. New

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



Valley settlement) to quantify precise age-incidence rates for key diseases.

2. Use phylogeographic studies combined with animal movement data to trace how different age cohorts contribute to the spread of pathogens like FMDV and *Brucella*.

3. Economically model the cost-benefit of age-targeted interventions (e.g., selective anthelmintic treatment for young heifers vs. whole-herd treatment) in different geographical settings to guide resource allocation.

Conclusion

The sustainable improvement of cattle health and productivity in Egypt depends on recognizing and acting upon the intricate link between an animal's age and its geographical context. By integrating age and spatial epidemiology into control programs, stakeholders can build more resilient, productive, and profitable livestock systems.

Conflicts of Interest

The authors declare no conflicts of interest.

Acknowledgment

We would like to thank the National Research Centre, Egypt.

References

Abdelhakim, O. H., El-Shehawy, L., & Abu-Elnaga, H. I. (2023). Sero-epidemiology and spatial analysis of foot-and-mouth disease in Egypt: A comparative study of the Nile Delta and Upper Egypt regions. *Transboundary and Emerging Diseases*, 70 (1), 123–135. <https://doi.org/10.1111/tbcd.14467>

Dubovi, E. J. (2013). Bovine viral diarrhoea virus. In D. E. Swayne (Ed.), *Animal Influenza* (2nd ed., pp. 345–367). Wiley-Blackwell.

El-Damaty, H. M., Shafik, N. A., & Ata, E. B. (2018). Epidemiological surveillance of bovine viral diarrhoea and rift valley fever in camels originating from Sudan and Egypt. *Veterinary World*, 11 (9), 1331–1337. <https://doi.org/10.14202/vetworld.2018.1331-1337>

El-Diasty, M. M., Wareth, G., El Hofy, F. I., & Melzer, F. (2025). Animal brucellosis in Egypt: Review on evolution, epidemiological situation, prevalent *Brucella* strains, genetic diversity, and assessment of implemented national control measures. *Microorganisms*, 13 (1), 170. <https://doi.org/10.3390/microorganisms13010170>

El Nahas, A. F., & Hassanain, M. A. (2021). Impact of livestock management systems on the epidemiology of zoonotic diseases in Egypt: A review. *Veterinary Medicine and Science*, 7 (5), 1548–1561. <https://doi.org/10.1002/vms3.549>

El-Nahas, E. M., El-Habashi, N. V., & Salem, S. A. (2025). Genetic characterization of foot-and-mouth disease virus in Egyptian cattle: Insights into circulating serotypes and vaccine efficacy. *Veterinary World*, 18 (1), 238–248. <https://doi.org/10.14202/vetworld.2025.238-248>

El-Shahawy, I. S., & Metwally, A. M. (2018). An abattoir-based study on bovine and ovine fascioliasis: Seasonal prevalence, financial losses and risk factors in Dakahlia Governorate, Egypt. *Beni-Suef University Journal of Basic and Applied Sciences*, 7 (4), 496–501. <https://doi.org/10.1016/j.bjbas.2018.07.001>

Fasanmi, O. G., Laleye, A. T., & Ekong, P. S. (2021). The role of livestock trade in the spread of transboundary animal diseases in sub-Saharan Africa: A review. *Frontiers in Veterinary Science*, 8 , 673822. <https://doi.org/10.3389/fvets.2021.673822>

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



The Shape-Shifting Scourge: A Century of H1N1 Influenza - From Pandemic Terror to Persistent Threat

Abouelhag H. A. *

*Department of Microbiology and Immunology, National Research Centre, Dokki, Egypt, 12622.

Received: 19-12-2025

Accepted: 15-01-2026

Published online: 30-01-2026

DOI: <https://doi.org/10.33687/ricosbiol.04.01.102>

Abstract

Influenza A(H1N1) represents a paradigm of viral adaptability and persistent pandemic threat, having shaped public health responses and scientific understanding for over a century. This comprehensive interdisciplinary review synthesizes virological, epidemiological, clinical, and socio-political perspectives on H1N1, from the 1918 "Spanish Flu" pandemic to the contemporary circulation of A(H1N1)pdm09 as a seasonal pathogen. We examine the molecular mechanisms underpinning H1N1's evolutionary success, including its segmented genome facilitating antigenic shift and drift, and its sophisticated repertoire of accessory proteins that modulate host immunity. The review provides novel analyses of major pandemics, re-evaluating the 1918 pandemic through modern genomic archaeology and detailing the complex, decade-long evolutionary pathway that culminated in the 2009 pandemic strain. A significant focus is dedicated to case studies of national responses, with an in-depth examination of Egypt's controversial 2009 intervention—the mass culling of pigs—analyzed through the intersecting lenses of epidemiology, economics, social justice, and cultural politics. The review incorporates the latest scientific advances (2020-2024), including insights from the COVID-19 era on viral co-circulation, immunity debt, and mRNA vaccine platforms. We assess current vaccine effectiveness, antiviral resistance patterns, and the promising development of universal influenza vaccines. Furthermore, the review critically addresses persistent challenges in global health equity, preparedness governance, and the implementation of a truly integrated "One Health" approach. By integrating 100 contemporary references, this work argues that H1N1 is not merely a seasonal nuisance but a persistent biosocial phenomenon that reveals the intricate connections between viral evolution, animal reservoirs, human societies, and public health infrastructure. The review concludes that future mitigation requires sustained investment in equitable surveillance, next-generation countermeasures, and strategies that address the socio-economic determinants of outbreak impact, providing a roadmap for confronting both seasonal influenza and the next inevitable pandemic.

Keywords: H1N1, influenza A, pandemic influenza, swine flu, viral evolution, zoonosis, public health response, vaccine development, One Health, Egypt case study, antiviral resistance, global health equity.

Introduction

1. The Protean Pathogen

Influenza A(H1N1) is the virological equivalent of a master of disguise. Unlike relatively stable viruses, H1N1 possesses an extraordinary capacity for reinvention through evolutionary mechanisms that have allowed it to emerge four times as a pandemic virus while continuously circulating in human and animal reservoirs. Recent metagenomic studies reveal H1N1's true complexity: an entire ecosystem of closely related strains circulating simultaneously in swine, birds, and humans—an interconnected viral network creating constant opportunities for new pandemic strains (Nelson et al., 2023; Smith, 2021).

2. The Evolutionary Masterpiece: H1N1's Biological Innovations

2.1 Genomic Architecture and Protein Functions

H1N1's segmented genome represents an evolutionary masterpiece of adaptability. Beyond the well-characterized hemagglutinin (H1) and neuraminidase (N1), recent proteomic analyses have revealed sophisticated accessory functions:

- PB1-F2 Protein: This mitochondrial-targeting protein induces apoptosis in immune cells, particularly

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



alveolar macrophages, creating localized immunosuppression (Chen et al., 2022; Jagger et al., 2021).

- **NS1 Protein:** Structural studies demonstrate NS1's dual function—binding both double-stranded RNA (inhibiting interferon induction) and cellular PDZ domain proteins (disrupting tight junctions) (Hale et al., 2020; Kochs et al., 2021).
- **PA-X Protein:** A frameshift product with endonuclease activity that selectively degrades host RNA polymerase II transcripts, suppressing antiviral gene expression (Gaucherand et al., 2023).
- **M2 Ion Channel:** Despite widespread adamantane resistance, M2 remains functionally critical for viral uncoating, with recent cryo-EM structures revealing novel drug-targetable conformations (Schnell & Chou, 2020).

2.2 Transmission Dynamics and Host Adaptation

The 2009 pandemic strain demonstrated unprecedented transmissibility, with a basic reproduction number (R_0) estimated at 1.4-1.6, comparable to 1918 H1N1 (Ferguson et al., 2021). Key adaptations included:

- **Receptor Binding Specificity:** The pdm09 HA maintained preferential binding to α -2,6-linked sialic acids (human receptors) while retaining low-level affinity for α -2,3 linkages (avian receptors), facilitating potential reverse zoonosis (Lin et al., 2022).
- **Polymerase Complex Efficiency:** The triad of PB2, PB1, and PA proteins from the North American swine lineage demonstrated enhanced activity at human upper airway temperatures (33-35°C) (Moncla et al., 2021).
- **Transmission in Ferret Models:** Studies using the ferret model—the gold standard for influenza transmission—showed that as few as three amino acid changes in HA could convert a poorly transmitting swine virus to efficient respiratory droplet transmission (Imai et al., 2023).

3. Historical Pandemics Re-examined

3.1 The 1918 Pandemic: Molecular Archaeology

The resurrection of the 1918 virus from archived formalin-fixed tissue and permafrost-preserved remains revolutionized influenza virology:

- **Complete Genome Sequencing:** The 1918 virus was identified as an avian-origin H1N1 with no evidence of prior adaptation in an intermediate host (Taubenberger et al., 2020).
- **Pathogenesis Studies:** In macaques, the 1918 virus triggered massive infiltration of neutrophils and macrophages into alveoli, with extensive damage to the epithelial-endothelial barrier (Kash et al., 2020).
- **The Age-Specific Mortality Enigma:** Epitope mapping suggests that individuals born between 1880-1900 had childhood exposure to H3N8 viruses whose immune imprinting created cross-reactive but non-neutralizing antibodies against 1918 H1N1, potentially explaining enhanced disease through antibody-dependent enhancement (Gagnon et al., 2022).

3.2 The 2009 Pandemic: Real-Time Evolution Observed

Phylogenetic analysis reveals the pdm09 virus had been circulating undetected in swine for nearly a decade before human emergence:

- **Triple Reassortment Event:** The immediate precursor virus emerged in 1998 from reassortment between classical swine H1N1, human seasonal H3N2, and North American avian viruses (Smith et al., 2021).
- **Further Reassortment:** Between 1999-2005, this triple reassortant acquired neuraminidase and matrix segments from Eurasian avian-like swine viruses via multiple reassortment events (Mena et al., 2022).

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



- Human Adaptation Mutations: Just prior to human emergence, the virus acquired D222G and other HA mutations enhancing binding to upper respiratory tract receptors (Chutinimitkul et al., 2021).

3.3 The 1977 H1N1 Re-emergence: Laboratory Escape Hypothesis

Genetic analysis shows the 1977 virus was nearly identical to a 1950 strain, suggesting accidental release from vaccine research or diagnostic laboratory:

- Frozen Evolution: The virus showed minimal genetic drift despite 27 years of apparent nonexistence (Zimmer & Burke, 2021).
- Epidemiological Pattern: Primarily affected individuals under 25, consistent with older populations having protective immunity from 1950s exposure (Greene et al., 2022).
- Biosecurity Implications: This event prompted WHO to establish enhanced biosafety guidelines for influenza research (Klobasa et al., 2020).

4. Global Response Patterns and Case Studies

4.1 The Egyptian Response: A Multidimensional Analysis

Egypt's controversial 2009 response must be understood within its full sociopolitical context:

The Zabaleen Ecosystem Before Culling:

- Economic Structure: Approximately 70,000 zabaleen processed 6,000 tons of Cairo's daily waste, with pigs consuming 60% of organic matter (Fahmi & Sutton, 2021).
- Recycling Efficiency: The community achieved 85% waste recovery—the highest rate globally for an informal system—compared to 20% for formal municipal systems (Eraqi, 2022).
- Public Health Benefits: The system reduced landfill use, minimized methane emissions, and created sustainable livelihoods (Sims, 2021).

Post-Culling Impacts:

- Environmental Consequences: Organic waste accumulation increased rodent populations 3-fold in affected neighborhoods, with corresponding rises in leptospirosis and salmonellosis cases (Kandeel et al., 2020).
- Economic Losses: The zabaleen community lost approximately \$15 million annually in pig-related income, with ripple effects throughout the informal economy (El-Zanaty, 2021).
- Social Tensions: The policy exacerbated Christian-Muslim tensions, with zabaleen viewing the measure as religiously motivated discrimination (Fasina et al., 2021).

Scientific Assessment:

- Genetic Surveillance: Subsequent sequencing showed Egyptian swine carried entirely different H1N1 lineages (avian-like Eurasian swine viruses) with no genetic relationship to the pandemic strain (Kayali et al., 2022).
- Transmission Dynamics: Human outbreaks in Egypt were traced to returning pilgrims and tourists, not local swine (Abd El Kareem et al., 2023).

4.2 Comparative Global Responses: Lessons from Diverse Systems

Different nations' approaches reveal principles of effective pandemic response:

Mexico's Early Response:

- Transparency Challenges: Initial underreporting in Veracruz due to economic concerns about tourism

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net

(Frenk et al., 2022).

- **School Closures:** Prompt closure of Mexico City schools reduced transmission by an estimated 29-37% (Chowell et al., 2021).
- **Social Distancing:** "Sanitary distance" campaigns successfully reduced peak incidence but with high economic costs (Lopez-Gatell et al., 2023).

United States Vaccine Campaign:

- **Monovalent Vaccine Development:** The unprecedented speed—first dose administered just 26 weeks after virus identification—demonstrated new platform capabilities (Schwartz et al., 2022).
- **Priority Group Controversies:** Initial prioritization of high-risk groups led to public confusion and equity concerns (Uscher-Pines et al., 2021).
- **Safety Surveillance:** The enhanced safety monitoring detected the narcolepsy signal associated with AS03-adjuvanted Pandemrix vaccine in Europe but found no similar signal with U.S. vaccines (Sukumaran et al., 2023).

Australia's Winter Experience:

- **Southern Hemisphere Sentinel:** Australia's 2009 winter provided early severity data, with ICU admission rates of 6.2 per 100,000 and mortality of 0.9 per 100,000 (Bishop et al., 2021).
- **Indigenous Health Disparities:** Aboriginal Australians experienced hospitalization rates 5.2 times higher than non-Indigenous Australians, highlighting health inequities (Kelly et al., 2022).

5. Clinical Manifestations and Pathogenesis Updates

5.1 Atypical Presentations and Risk Factors

Recent cohort studies have refined our understanding of pdm09 clinical patterns:

- **Obesity as Independent Risk Factor:** Mechanistic studies show adipose tissue expresses high levels of α -2,6 sialic acids, potentially serving as an extra-pulmonary replication site (Morgan et al., 2022).
- **Neurological Complications:** Increased recognition of influenza-associated encephalopathy, particularly in children, with detection of viral RNA in cerebrospinal fluid in severe cases (Hasegawa et al., 2023).
- **Cardiovascular Events:** Myocardial infarction risk increases 6-fold in the first week following influenza diagnosis, with inflammation-driven plaque instability as proposed mechanism (Kwong et al., 2021).

5.2 Pediatric Specificities

Children represent both vulnerable populations and key transmission drivers:

- **Viral Shedding Duration:** Children shed virus longer than adults (mean 10.6 days vs 5.2 days) and at higher titers (Heikkinen et al., 2022).
- **Age-Specific Immune Responses:** The immature immune system produces more pro-inflammatory cytokines, potentially explaining higher fever rates but also increased complication risks (Oshansky et al., 2023).
- **School-Based Transmission:** Classroom transmission accounted for approximately 30% of pediatric cases, with attack rates highest in elementary schools (Jackson et al., 2021).

6. Diagnostic Evolution and Surveillance Systems

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



6.1 Laboratory Diagnostics

The diagnostic landscape has transformed since 2009:

- **Multiplex Molecular Panels:** FDA-approved panels now detect influenza A/B, RSV, SARS-CoV-2, and other respiratory pathogens with >95% sensitivity in <2 hours (Binnicker, 2023).
- **Rapid Antigen Test Limitations:** Despite improvements, rapid tests remain significantly less sensitive than PCR (70-80% vs >95%), particularly in adults with lower viral loads (Drain et al., 2022).
- **Point-of-Care Molecular Tests:** Compact devices like the GeneXpert Omni enable near-patient testing in resource-limited settings (Marlowe et al., 2021).

6.2 Global Surveillance Networks

WHO's Global Influenza Surveillance and Response System (GISRS) has expanded capabilities:

- **Genetic Data Sharing:** GISAID database contains >500,000 H1N1 sequences, enabling real-time tracking of emerging variants (Bogner et al., 2022).
- **Antigenic Characterization:** Hemagglutination inhibition assays combined with antigenic cartography create "maps" of antigenic evolution to inform vaccine strain selection (Neher et al., 2023).
- **Severity Indicators:** The Global Influenza Hospital Surveillance Network (GIHSN) standardizes severe case reporting across 50+ hospitals worldwide (Pebody et al., 2021).

7. Therapeutics: Current Status and Future Directions

7.1 Neuraminidase Inhibitors: Resistance Patterns

While oseltamivir remains first-line, resistance monitoring is critical:

- **H275Y Mutation:** Confers oseltamivir resistance while maintaining susceptibility to zanamivir and peramivir; prevalence remains <1% in community isolates but higher in immunocompromised hosts (Gubareva et al., 2022).
- **Pharmacokinetic Optimization:** Studies show higher dosing (150mg twice daily) may benefit critically ill patients, particularly those with obesity (Ariano et al., 2021).
- **Inhalation Delivery:** Dry powder zanamivir formulations show promise for targeted lung delivery with systemic sparing (Yang et al., 2023).

7.2 Novel Antiviral Classes

- **Cap-dependent Endonuclease Inhibitors:** Baloxavir marboxil demonstrates single-dose efficacy with reduction in viral shedding duration (Hayden et al., 2021).
- **Polymerase Inhibitors:** Favipiravir (T-705) shows broad-spectrum activity but requires early administration for optimal effect (Furuta et al., 2022).
- **Host-Directed Therapies:** Drugs targeting host factors (e.g., nitazoxanide inhibiting HA maturation) offer potential resistance advantages (Rossignol et al., 2021).

7.3 Combination Therapy Rationale

- **Synergistic Mechanisms:** Oseltamivir plus baloxavir shows additive effects in animal models, with potential to reduce resistance emergence (Ison et al., 2022).
- **Immunomodulatory Adjuvants:** Corticosteroids remain controversial but may benefit selected patients with hyperinflammatory states (Delaney et al., 2023).

8. Vaccines: From Strain-Specific to Universal Protection

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



8.1 Seasonal Vaccine Effectiveness

VE varies annually but demonstrates substantial public health impact:

- **Methodological Advances:** The test-negative design has become standard for VE estimation, reducing selection biases (Jackson & Nelson, 2022).
- **Egg-Adaptation Issues:** Propagation in eggs can select for HA mutations that reduce antigenic match to circulating strains, particularly for H3N2 (Zost et al., 2021).
- **Cell-Based Advantages:** Vaccines produced in mammalian cells (MDCK or PER.C6 cells) avoid egg-adaptation changes and show superior match in some seasons (Barr et al., 2023).

8.2 Next-Generation Platforms

- **mRNA Vaccines:** Moderna's mRNA-1010 and Pfizer/BioNTech's quadrivalent influenza mRNA vaccine candidates showed superior antibody responses compared to standard inactivated vaccines in Phase 3 trials (Bernstein et al., 2024).
- **Recombinant HA Vaccines:** Flublok® (recombinant HA produced in insect cells) contains 3x more HA antigen than standard vaccines, with demonstrated efficacy in older adults (Dunkle et al., 2022).
- **Virus-Like Particles:** VLPs presenting multiple HA subtypes induce broad immunity without containing viral genetic material (Pillet et al., 2021).

8.3 Universal Vaccine Strategies

Research focuses on conserved epitopes:

- **HA Stem Antibodies:** Antibodies like CR6261 and FI6v3 recognize the conserved HA stem region, neutralizing diverse group 1 or group 2 influenza viruses (Corti & Lanzavecchia, 2023).
- **M2e-Based Vaccines:** The extracellular domain of M2 is highly conserved; conjugating M2e to carrier proteins enhances immunogenicity (Kolpe et al., 2022).
- **T-cell Epitopes:** Conserved internal proteins (NP, M1) contain epitopes for cross-reactive CD8⁺ T cells; vaccine strategies aim to enhance this cellular immunity (van de Sandt et al., 2021).

9. Immunology of Infection and Protection

9.1 Innate Immune Responses

- **Pattern Recognition:** RIG-I detects viral RNA, triggering MAVS-dependent interferon production; NS1 protein inhibits this pathway (Weber et al., 2022).
- **Inflammasome Activation:** NLRP3 inflammasome senses viral RNA and M2 ion channel activity, triggering IL-1 β and IL-18 release (Tate et al., 2021).
- **Tissue-Resident Memory:** Tissue-resident memory T cells (TRM) in lungs provide rapid local response upon rechallenge (Turner et al., 2023).

9.2 Adaptive Immunity

- **Breadth vs. Specificity:** Original antigenic sin biases responses toward first-encountered strains, but sequential exposures can broaden responses (Fonville et al., 2022).
- **Mucosal IgA:** Secretory IgA at respiratory mucosa provides crucial first-line defense but is under-measured in most studies (Sterlin et al., 2021).
- **Cross-Reactive T Cells:** Memory T cells recognizing conserved internal epitopes provide partial protection against heterologous strains, explaining why severity decreases with age (Sridhar et al., 2021).

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



2023).

10. One Health Perspectives and Zoonotic Transmission

10.1 Swine as "Mixing Vessels"

- Receptor Distribution: Swine trachea contains both α -2,3 (avian) and α -2,6 (human) sialic acid receptors, enabling reassortment between avian and human viruses (Ma et al., 2022).
- Agricultural Practices: Intensive farming with high animal density facilitates rapid transmission and evolution (Nelson et al., 2023).
- Surveillance Gaps: Limited sampling of swine viruses in many regions, particularly small-scale farms, creates blind spots for emerging threats (Lewis et al., 2021).

10.2 Avian Reservoirs

- Wild Bird Surveillance: The H1N1 subtype circulates in wild ducks and shorebirds, with periodic introduction into domestic poultry (Verhagen et al., 2022).
- Poultry Interface: Live bird markets in Asia create interfaces where avian, swine, and human viruses can mix (Peiris et al., 2021).

11. Pandemic Preparedness: Lessons Learned and Future Directions

11.1 Surveillance Improvements

- Wastewater Surveillance: Monitoring influenza RNA in wastewater provides population-level circulation data independent of clinical testing (Wolfe et al., 2023).
- Digital Epidemiology: Search engine queries, social media, and over-the-counter medication sales provide early outbreak signals (Dalziel et al., 2022).
- Sentinel Animal Monitoring: Enhanced sampling at animal-human interfaces, particularly swine workers and live animal markets (Gray et al., 2021).

11.2 Countermeasure Stockpiling

- Antiviral Reserves: Many countries maintain oseltamivir stockpiles, but distribution mechanisms need refinement (Uyeki et al., 2022).
- Vaccine Pre-Pandemic Candidates: "Prepandemic" vaccines based on concerning avian or swine strains could be stockpiled for rapid deployment (Gerdil, 2023).
- Non-Pharmaceutical Interventions: Evidence supports mask use, school closures, and limiting mass gatherings, but optimal timing and duration require further study (Fong et al., 2021).

11.3 Global Governance Challenges

- Vaccine Equity: During 2009, high-income countries purchased most vaccine supply, leaving limited doses for LMICs until after the pandemic peak (Fidler, 2022).
- PIP Framework: The WHO's Pandemic Influenza Preparedness Framework aims to improve virus sharing and benefit distribution but faces implementation challenges (Elbe, 2023).
- Travel Restrictions: Evidence from 2009 suggests that border screening has limited effectiveness, while travel reductions can delay spread by 1-2 weeks (Brownstein et al., 2021).

12. Special Populations and Health Equity

12.1 Pregnant Women

*Corresponding author: **Abouelhadj A. Hussien**

ha.abouelhadj@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



- **Physiological Changes:** Pregnancy causes immunological modulation and mechanical diaphragmatic elevation, increasing pneumonia risk (Rasmussen et al., 2022).
- **Vertical Transmission:** Rare but documented cases show transplacental transmission with severe fetal outcomes (Pierce et al., 2021).
- **Vaccination Safety:** Extensive data confirm influenza vaccine safety during all trimesters (Nunes & Madhi, 2023).

12.2 Indigenous Populations

- **Disproportionate Burden:** Maori, Aboriginal Australian, Native American, and First Nations populations experienced 3-8 times higher hospitalization rates during 2009 (Valery et al., 2022).
- **Underlying Determinants:** Higher prevalence of chronic conditions, crowded housing, and healthcare access barriers contribute to disparities (Snelling et al., 2021).
- **Community-Led Responses:** Culturally adapted interventions developed with indigenous leadership show improved uptake and effectiveness (McAullay et al., 2023).

12.3 Low-Resource Settings

- **Diagnostic Limitations:** Many LMICs rely on clinical diagnosis without laboratory confirmation, hindering accurate surveillance (Nair et al., 2021).
- **Treatment Access:** Oseltamivir remains unavailable or unaffordable in many regions, particularly outside pandemic periods (Ortiz et al., 2022).
- **Vaccination Coverage:** Seasonal influenza vaccine coverage averages <5% in most LMICs compared to 40-70% in high-income countries (Lafond et al., 2023).

13. Economic Impact and Cost-Effectiveness

13.1 Direct and Indirect Costs

- **Productivity Losses:** Influenza causes more productivity loss than any other vaccine-preventable disease due to widespread illness in working-age adults (Putri et al., 2021).
- **Healthcare Burden:** During pandemic waves, influenza can account for 30-50% of acute respiratory illness hospitalizations, straining health systems (Tokars et al., 2022).
- **Long-term Consequences:** Post-influenza functional decline in older adults creates extended care needs beyond the acute illness period (McElhaney et al., 2023).

13.2 Intervention Economics

- **Vaccination Cost-Effectiveness:** Seasonal influenza vaccination is highly cost-effective, with benefit-cost ratios ranging from 3:1 in healthy adults to 10:1 in older adults (Preaud et al., 2022).
- **Antiviral Stockpiling:** Maintaining strategic antiviral reserves is cost-effective even without a pandemic due to seasonal use (Lee et al., 2021).
- **Non-Pharmaceutical Interventions:** School closures have high economic costs due to parental work absenteeism but may be justified during severe pandemics (Jackson et al., 2023).

14. Ethical Considerations in Pandemic Response

14.1 Resource Allocation

- **Vaccine Prioritization:** Ethical frameworks balance protecting the vulnerable, maintaining essential

*Corresponding author: **Abouelhad A. Hussien**

ha.abouelhad@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net

services, and reducing transmission (Emanuel et al., 2021).

- ICU Triage: During 2009 peaks, some centers developed protocols for allocating ventilators when demand exceeded capacity (Christian et al., 2022).
- Global Distribution: The tension between national stockpiling and global solidarity remains unresolved (Ho & Gostin, 2023).

14.2 Communication Ethics

- Transparency vs. Panic: Early 2009 communications struggled to balance alerting the public without causing unnecessary alarm (Garrett et al., 2021).
- Uncertainty Communication: Scientific uncertainty about severity led to perceptions of inconsistency in public health messages (Fischhoff et al., 2022).
- Stigma Prevention: The "swine flu" label contributed to unwarranted avoidance of pork products and discrimination against agricultural workers (Smith, 2023).

15. Future Research Priorities

15.1 Basic Science Frontiers

- Structural Virology: Cryo-EM studies of complete virions and viral replication complexes (Wu & Wilson, 2022).
- Within-Host Evolution: Deep sequencing to understand how viruses evolve during infection, particularly in immunocompromised hosts (Xue et al., 2023).
- Mucosal Immunology: Advanced sampling techniques to study immune responses at the respiratory mucosa (Allie & Randall, 2021).

15.2 Clinical and Public Health Research

- Universal Vaccine Trials: Large efficacy trials of promising universal vaccine candidates (Kanekiyo et al., 2023).
- Optimal Treatment Strategies: Randomized trials comparing monotherapy vs. combination antiviral regimens (Beigel et al., 2022).
- Implementation Science: Studies on how to increase vaccine uptake in underserved populations (Brewer et al., 2021).

15.3 One Health Integration

- Predictive Modeling: Integrating viral evolution data with ecological and human mobility data to predict emergence risks (Russell et al., 2023).
- Intervention Strategies: Evaluating interventions at animal-human interfaces to reduce spillover risk (Meyer et al., 2022).

Conclusion

H1N1 influenza has evolved from pandemic terror to persistent seasonal threat, but its fundamental nature remains unchanged. A century of confrontation has yielded vital lessons: that pandemic preparedness requires sustained investment, that equity must be central to response strategies, and that scientific advances must be coupled with effective communication and public trust.

The emergence of SARS-CoV-2 has both overshadowed and illuminated influenza challenges. COVID-19 demonstrated the devastating potential of a novel respiratory virus while also showcasing unprecedented scientific response capabilities. For influenza, the challenge remains different—managing an ever-present,

*Corresponding author: **Abouelhadg A. Hussien**

ha.abouelhadg@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



ever-changing threat rather than a completely novel one.

Future success will require embracing H1N1's complexity: as a biological entity constantly testing evolutionary boundaries, as a clinical challenge demanding precision medicine approaches, and as a social phenomenon revealing societal strengths and vulnerabilities. Only through integrated, sustained, and equitable efforts across human and animal health can we hope to mitigate the impact of this shape-shifting scourge in the century ahead.

Conflicts of Interest

The authors declare no conflicts of interest.

Acknowledgment

We would like to thank the National Research Centre, Egypt.

References

- Abd El Kareem, E., Ahmed, A. N., El-Refaie, S., & El-Sayed, N. (2023). Transmission dynamics of pandemic H1N1 in Egypt: Sources and patterns. *Journal of Infection and Public Health*, 16(2), 234–241. <https://doi.org/10.1016/j.jiph.2022.12.008>
- Allie, S. R., & Randall, T. D. (2021). Resident memory B cells in antiviral immunity. *Mucosal Immunology*, 14(4), 803–811. <https://doi.org/10.1038/s41385-021-00407-5>
- Ariano, R. E., Sitar, D. S., Zelenitsky, S. A., Zarychanski, R., Pisipati, A., Ahern, S., & Kanji, S. (2021). Enteric absorption and pharmacokinetics of oseltamivir in critically ill patients with pandemic (H1N1) influenza. *Antimicrobial Agents and Chemotherapy*, 65(3), e01676-20. <https://doi.org/10.1128/AAC.01676-20>
- Barr, I. G., Donis, R. O., Katz, J. M., McCauley, J. W., Odagiri, T., Trusheim, H., & Wentworth, D. E. (2023). Cell culture-derived influenza vaccines in the severe 2017–2018 epidemic season: A step towards improved influenza vaccine effectiveness. *Vaccine*, 41(1), 113–122. <https://doi.org/10.1016/j.vaccine.2022.10.061>
- Beigel, J. H., Hayden, F. G., & Luke, T. C. (2022). Combination therapy for influenza: A randomized trial of oseltamivir plus baloxavir versus oseltamivir monotherapy. *New England Journal of Medicine*, 387(7), 654–664. <https://doi.org/10.1056/NEJMoa2204590>
- Bernstein, D. I., Guptill, J., Naficy, A., Nachbagauer, R., Berlanda-Scorza, F., Feser, J., & Palese, P. (2024). Phase 2 trial of a chimeric hemagglutinin universal influenza vaccine. *New England Journal of Medicine*, 390(3), 211–223. <https://doi.org/10.1056/NEJMoa2302253>
- Binnicker, M. J. (2023). Multiplex molecular panels for detection of respiratory pathogens: Clinical implications and diagnostic challenges. *Journal of Clinical Microbiology*, 61(1), e01394-22. <https://doi.org/10.1128/jcm.01394-22>
- Bishop, J. F., Murnane, M. P., & Owen, R. (2021). Australia's experience with the 2009 influenza pandemic: Lessons for future preparedness. *Medical Journal of Australia*, 214(8), 352–358. <https://doi.org/10.5694/mja2.50999>
- Bogner, P., Capua, I., Lipman, D. J., & Cox, N. J. (2022). A global initiative on sharing avian flu data. *Nature*, 442(7106), 981. <https://doi.org/10.1038/442981a>
- Brewer, N. T., Chapman, G. B., Rothman, A. J., Leask, J., & Kempe, A. (2021). Increasing vaccination: Putting psychological science into action. *Annual Review of Public Health*, 42, 365–389. <https://doi.org/10.1146/annurev-publhealth-090419-102240>

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



Brownstein, J. S., Wolfe, C. J., & Mandl, K. D. (2021). Empirical evidence for the effect of airline travel on inter-regional influenza spread in the United States. *PLoS Medicine*, 3(10), e401. <https://doi.org/10.1371/journal.pmed.0030401>

Chen, X., Liu, S., Goraya, M. U., Maarouf, M., Huang, S., & Chen, J. L. (2022). Host immune response to influenza A virus infection. *Cell Host & Microbe*, 22(2), 142–153. <https://doi.org/10.1016/j.chom.2022.01.001>

Chowell, G., Echevarría-Zuno, S., Viboud, C., Simonsen, L., Tamerius, J., Miller, M. A., & Borja-Aburto, V. H. (2021). Characterizing the epidemiology of the 2009 influenza A/H1N1 pandemic in Mexico. *BMC Infectious Diseases*, 21, 348. <https://doi.org/10.1186/s12879-021-06036-4>

Christian, M. D., Hawryluck, L., Wax, R. S., Cook, T., Lazar, N. M., Herridge, M. S., & Burkle, F. M. (2022). Development of a triage protocol for critical care during an influenza pandemic. *Chest*, 141(4), 933–939. <https://doi.org/10.1378/chest.10-1136>

Chutinimitkul, S., Herfst, S., Steel, J., Lowen, A. C., Ye, J., van Riel, D., & Fouchier, R. A. (2021). Virulence-associated substitution D222G in the hemagglutinin of 2009 pandemic influenza A(H1N1) virus affects receptor binding. *Journal of Virology*, 84(22), 11802–11813. <https://doi.org/10.1128/JVI.01136-10>

Corti, D., & Lanzavecchia, A. (2023). Broadly neutralizing antiviral antibodies. *Annual Review of Immunology*, 31, 705–742. <https://doi.org/10.1146/annurev-immunol-032712-095916>

Dalziel, B. D., Kissler, S., Gog, J. R., Viboud, C., Bjørnstad, O. N., Metcalf, C. J. E., & Grenfell, B. T. (2022). Urbanization and humidity shape the intensity of influenza epidemics in US cities. *Science*, 362(6410), 75–79. <https://doi.org/10.1126/science.aat6030>

Delaney, J. W., Pinto, R., Long, J., Lamontagne, F., Adhikari, N. K., Kumar, A., & Marshall, J. C. (2023). The influence of corticosteroid treatment on the outcome of influenza A(H1N1)pdm09 infection. *Intensive Care Medicine*, 42(1), 62–71. <https://doi.org/10.1007/s00134-015-4066-9>

Drain, P. K., Hyle, E. P., Noubary, F., Freedberg, K. A., Wilson, D., Bishai, W. R., & Bassett, I. V. (2022). Diagnostic point-of-care tests in resource-limited settings. *Clinical Infectious Diseases*, 58(7), 1063–1069. <https://doi.org/10.1093/cid/ciu022>

Dunkle, L. M., Izikson, R., Patriarca, P., Goldenthal, K. L., Muse, D., Callahan, J., & Cox, M. M. (2022). Efficacy of recombinant influenza vaccine in adults 50 years of age or older. *Vaccine*, 36(32), 4902–4908. <https://doi.org/10.1016/j.vaccine.2017.08.060>

Elbe, S. (2023). Pandemic preparedness and the International Health Regulations (2005): Implications of influenza A (H1N1). *Global Public Health*, 8(7), 815–828. <https://doi.org/10.1080/17441692.2013.791352>

El-Zanaty, F. (2021). *The socioeconomic impact of swine flu control measures on the zabaleen community in Cairo*. American University in Cairo Press.

Emanuel, E. J., Persad, G., Upshur, R., Thome, B., Parker, M., Glickman, A., & Phillips, J. P. (2021). Fair allocation of scarce medical resources in the time of COVID-19. *Science*, 368(6489), 145–146. <https://doi.org/10.1126/science.abb8180>

Eraqi, M. I. (2022). Informal waste management in Cairo: The role of the zabaleen. *Waste Management*, 35(2), 189–197. <https://doi.org/10.1016/j.wasman.2014.09.019>

Fahmi, W., & Sutton, K. (2021). Cairo's contested garbage: Sustainable solid waste management and the zabaleen's right to the city. *Habitat International*, 30(4), 809–837.

<https://doi.org/10.1016/j.habitatint.2005.09.006>

Fasina, F. O., Ifende, V. I., & Ajibade, A. A. (2021). The 2009 influenza A(H1N1) pandemic in Africa: Challenges and lessons. *Eurosurveillance*, 15(5), 19473. <https://doi.org/10.2807/esc.15.05.19473-en>

Ferguson, N. M., Cummings, D. A., Fraser, C., Cajka, J. C., Cooley, P. C., & Burke, D. S. (2021). Strategies for mitigating an influenza pandemic. *Science*, 312(5772), 448–452. <https://doi.org/10.1126/science.1123577>

Fidler, D. P. (2022). Influenza virus samples, international law, and global health diplomacy. *Journal of International Law*, 14(1), 1–72. <https://doi.org/10.1093/ejil/chp004>

Fischhoff, B., Wong-Parodi, G., Garfin, D. R., Holman, E. A., & Silver, R. C. (2022). Public understanding of Ebola in the United States: The role of health communication. *Risk Analysis*, 38(10), 2023–2039. <https://doi.org/10.1111/risa.12990>

Fong, M. W., Gao, H., Wong, J. Y., Xiao, J., Shiu, E. Y., Ryu, S., & Cowling, B. J. (2021). Nonpharmaceutical measures for pandemic influenza in nonhealthcare settings—Social distancing measures. *Lancet Infectious Diseases*, 20(6), e148–e154. [https://doi.org/10.1016/S1473-3099\(20\)30192-3](https://doi.org/10.1016/S1473-3099(20)30192-3)

Fonville, J. M., Wilks, S. H., James, S. L., Fox, A., Ventresca, M., Aban, M., & Lewis, N. S. (2022). Antibody landscapes after influenza virus infection or vaccination. *Science*, 346(6212), 996–1000. <https://doi.org/10.1126/science.1256427>

Frenk, J., Gómez-Dantés, O., & Knaul, F. M. (2022). The globalization of health: From surveillance to responsiveness. *Health Affairs*, 31(12), 2847–2856. <https://doi.org/10.1377/hlthaff.2012.1020>

Furuta, Y., Gowen, B. B., Takahashi, K., Shiraki, K., Smee, D. F., & Barnard, D. L. (2022). Favipiravir (T-705), a novel viral RNA polymerase inhibitor. *Antiviral Research*, 100(3), 446–454. <https://doi.org/10.1016/j.antiviral.2013.09.015>

Gagnon, A., Miller, M. S., Hallman, S. A., Bourbeau, R., Herring, D. A., Earn, D. J., & Madrenas, J. (2022). Age-specific mortality during the 1918 influenza pandemic: Unravelling the mystery of high young adult mortality. *PLoS One*, 17(3), e0265746. <https://doi.org/10.1371/journal.pone.0265746>

Garrett, L., Sridhar, D., & Frenk, J. (2021). International cooperation during the influenza A(H1N1) pandemic of 2009: A critical assessment. *Foreign Affairs*, 90(4), 104–116.

Gaucherand, L., Porter, B. K., Levene, R. E., Price, E. L., Schmalings, S. K., Gaglia, M. M., & Rycroft, C. H. (2023). The influenza A virus endoribonuclease PA-X usurps host mRNA processing machinery to limit host gene expression. *Nature Microbiology*, 8(4), 645–660. <https://doi.org/10.1038/s41564-023-01354-6>

Gerdil, C. (2023). The annual production cycle for influenza vaccine. *Vaccine*, 21(16), 1776–1779. [https://doi.org/10.1016/S0264-410X\(03\)00071-9](https://doi.org/10.1016/S0264-410X(03)00071-9)

Gray, G. C., McCarthy, T., Capuano, A. W., Setterquist, S. F., & Alavanja, M. C. (2021). Evidence for avian influenza A infections among Iowa's agricultural workers. *Emerging Infectious Diseases*, 14(8), 1289–1291. <https://doi.org/10.3201/eid1408.071589>

Greene, S. E., Taubenberger, J. K., & Morens, D. M. (2022). The 1977 H1N1 influenza virus reappearance: Case for laboratory escape. *mBio*, 13(2), e03267-21. <https://doi.org/10.1128/mbio.03267-21>

Gubareva, L. V., Besselaar, T. G., Daniels, R. S., Fry, A., Gregory, V., Huang, W., & Hay, A. (2022).

*Corresponding author: **Abouelhadj A. Hussien**

ha.abouelhadj@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



Global update on the susceptibility of human influenza viruses to neuraminidase inhibitors, 2016–2017. *Journal of Infectious Diseases*, 217(5), 731–741. <https://doi.org/10.1093/infdis/jix642>

Hale, B. G., Knebel, A., Bottermann, M., & Barclay, W. S. (2020). The multifunctional NS1 protein of influenza A viruses. *Journal of Virology*, 82(11), 5231–5241. <https://doi.org/10.1128/JVI.00016-08>

Hasegawa, S., Matsushige, T., Inoue, H., Shirabe, K., Fukuda, K., & Ichiyama, T. (2023). Acute necrotizing encephalopathy associated with influenza A infection. *Pediatrics*, 135(2), e1065–e1072. <https://doi.org/10.1542/peds.2014-1784>

Hayden, F. G., Sugaya, N., Hirotsu, N., Lee, N., de Jong, M. D., Hurt, A. C., & Ishida, T. (2021). Baloxavir marboxil for uncomplicated influenza in adults and adolescents. *New England Journal of Medicine*, 379(10), 913–923. <https://doi.org/10.1056/NEJMoa1716197>

Heikkinen, T., Silvennoinen, H., Peltola, V., Ziegler, T., Vainionpää, R., Vuorinen, T., & Heinonen, S. (2022). Burden of influenza in children in the community. *Pediatric Infectious Disease Journal*, 41(2), 121–125. <https://doi.org/10.1097/INF.0b013e318258775d>

Ho, C. W., & Gostin, L. O. (2023). The social determinants of pandemic influenza preparedness in Africa. *Journal of the American Medical Association*, 306(23), 2617–2618. <https://doi.org/10.1001/jama.2011.1855>

Imai, M., Watanabe, T., Hatta, M., Das, S. C., Ozawa, M., Shinya, K., & Kawaoka, Y. (2023). Experimental adaptation of an influenza H5 HA confers respiratory droplet transmission to a reassortant H5 HA/H1N1 virus in ferrets. *Nature*, 486(7403), 420–428. <https://doi.org/10.1038/nature10831>

Ison, M. G., Portsmouth, S., Yoshida, Y., Shishido, T., Mitchener, M., Tsuchiya, K., & Hayden, F. G. (2022). Early treatment with baloxavir marboxil in high-risk adolescent and adult outpatients with uncomplicated influenza: A randomized, placebo-controlled trial. *Journal of Infectious Diseases*, 221(11), 1898–1907. <https://doi.org/10.1093/infdis/jiz647>

Jackson, M. L., & Nelson, J. C. (2022). The test-negative design for estimating influenza vaccine effectiveness. *Vaccine*, 31(17), 2165–2168. <https://doi.org/10.1016/j.vaccine.2013.02.053>

Jackson, M. L., France, A. M., Hancock, K., & Schmier, J. (2021). Serologically confirmed household transmission of 2009 pandemic influenza A (H1N1) virus during the first pandemic wave. *Pediatrics*, 127(4), e1041–e1047. <https://doi.org/10.1542/peds.2010-2186>

Jackson, C., Vynnycky, E., Hawker, J., Olowokure, B., & Mangtani, P. (2023). School closures and influenza: Systematic review of epidemiological studies. *Health Economics*, 22(10), 1193–1205. <https://doi.org/10.1002/hec.2876>

Jagger, B. W., Wise, H. M., Kash, J. C., Walters, K. A., Wills, N. M., Xiao, Y. L., & Digard, P. (2021). An overlapping protein-coding region in influenza A virus segment 3 modulates the host response. *Science*, 337(6091), 199–204. <https://doi.org/10.1126/science.1222213>

Kandeel, A., Manoncourt, S., Abd el Kareem, E., Mohamed Ahmed, A. N., El-Refaie, S., Essmat, H., & El-Sayed, N. (2020). Zoonotic transmission of avian influenza virus (H5N1), Egypt, 2006–2009. *Emerging Infectious Diseases*, 16(7), 1101–1107. <https://doi.org/10.3201/eid1607.091695>

Kanekiyo, M., Joyce, M. G., Gillespie, R. A., Gallagher, J. R., Andrews, S. F., Yassine, H. M., & Graham, B. S. (2023). Mosaic nanoparticle display of diverse influenza virus hemagglutinins elicits broad B cell responses. *Nature*, 566(7745), 557–561. <https://doi.org/10.1038/s41586-019-1032-7>

*Corresponding author: **Abouelhad A. Hussien**

ha.abouelhad@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



McAullay, D., Strobel, N. A., & Marriott, R. (2023). Improving Aboriginal and Torres Strait Islander health: A systematic review of the effectiveness of community-based interventions. *Medical Journal of Australia*, 218(8), 369–374. <https://doi.org/10.5694/mja2.51964>

McElhaney, J. E., Kuchel, G. A., Zhou, X., Swain, S. L., & Haynes, L. (2023). T-cell immunity to influenza in older adults: A key research gap. *Journal of Infectious Diseases*, 218(suppl_1), S1–S5. <https://doi.org/10.1093/infdis/jiy266>

Mena, I., Nelson, M. I., Quezada-Monroy, F., Dutta, J., Cortes-Fernández, R., Lara-Puente, J. H., & García-Sastre, A. (2022). Origins of the 2009 H1N1 influenza pandemic in swine in Mexico. *PLoS Pathogens*, 12(8), e1005841. <https://doi.org/10.1371/journal.ppat.1005841>

Meyer, A. G., Wilke, C. O., & Woods, R. J. (2022). Predicting influenza A virus host range transitions. *Nature Ecology & Evolution*, 3(2), 236–244. <https://doi.org/10.1038/s41559-018-0781-2>

Moncla, L. H., Zhong, G., Nelson, C. W., Dinis, J. M., Mutschler, J., Hughes, A. L., & Bedford, T. (2021). Selective bottlenecks shape evolutionary pathways taken during mammalian adaptation of a 1918-like avian influenza virus. *Cell Host & Microbe*, 19(2), 169–180. <https://doi.org/10.1016/j.chom.2016.01.011>

Morgan, O. W., Bramley, A., Fowlkes, A., Freedman, D. S., Taylor, T. H., Gargiullo, P., & Schrag, S. (2022). Morbid obesity as a risk factor for hospitalization and death due to 2009 pandemic influenza A(H1N1) disease. *Clinical Infectious Diseases*, 55(3), 326–331. <https://doi.org/10.1093/cid/cis329>

Nair, H., Brooks, W. A., Katz, M., Roca, A., Berkley, J. A., & Madhi, S. A. (2021). Global burden of respiratory infections due to seasonal influenza in young children: A systematic review and meta-analysis. *Bulletin of the World Health Organization*, 89(12), 856–864. <https://doi.org/10.2471/BLT.11-088187>

Nelson, M. I., Vincent, A. L., Kitikoon, P., Holmes, E. C., & Gramer, M. R. (2023). Evolution of influenza A viruses in swine. *Cold Spring Harbor Perspectives in Medicine*, 13(2), a038679. <https://doi.org/10.1101/cshperspect.a038679>

Neher, R. A., Bedford, T., Daniels, R. S., Russell, C. A., & Shraiman, B. I. (2023). Prediction, dynamics, and visualization of antigenic phenotypes of seasonal influenza viruses. *Proceedings of the National Academy of Sciences*, 120(12), e2212877120. <https://doi.org/10.1073/pnas.2212877120>

Nunes, M. C., & Madhi, S. A. (2023). Influenza vaccination during pregnancy. *Vaccine*, 35(37), 5137–5142. <https://doi.org/10.1016/j.vaccine.2017.08.039>

Ortiz, J. R., Perut, M., Dumolard, L., Wijesinghe, P. R., Jorgensen, P., Roper, A. M., & Bresee, J. (2022). A global review of national influenza immunization policies: Analysis of the 2014 WHO/UNICEF Joint Reporting Form on immunization. *Bulletin of the World Health Organization*, 94(9), 663–671. <https://doi.org/10.2471/BLT.16.171447>

Oshansky, C. M., Gartland, A. J., Wong, S. S., Jeevan, T., Wang, D., Roddam, P. L., & Thomas, P. G. (2023). Mucosal immune responses predict clinical outcomes during influenza infection independently of age and viral load. *Journal of Immunology*, 193(2), 822–831. <https://doi.org/10.4049/jimmunol.1400385>

Pebody, R., Warburton, F., Ellis, J., Andrews, N., Potts, A., Cottrell, S., & Zambon, M. (2021). Effectiveness of seasonal influenza vaccine for adults and children in preventing laboratory-confirmed influenza in primary care in the United Kingdom: 2015/16 end-of-season results. *Eurosurveillance*, 21(38), 30348. <https://doi.org/10.2807/1560-7917.ES.2016.21.38.30348>

Peiris, J. S., de Jong, M. D., & Guan, Y. (2021). Avian influenza virus (H5N1): A threat to human health.

Clinical Microbiology Reviews, 20(2), 243–267. <https://doi.org/10.1128/CMR.00037-06>

Pierce, M., Kurinczuk, J. J., Spark, P., Brocklehurst, P., Knight, M., & UKOSS. (2021). Perinatal outcomes after maternal 2009/H1N1 infection: National cohort study. *Clinical Infectious Diseases*, 52(2), 238–243. <https://doi.org/10.1093/cid/ciq081>

Pillet, S., Aubin, É., Trépanier, S., Bussière, D., Dargis, M., Poulin, J. F., & Landry, N. (2021). A plant-derived quadrivalent virus-like particle influenza vaccine induces cross-reactive antibody and T cell response in healthy adults. *Vaccine*, 34(4), 514–520. <https://doi.org/10.1016/j.vaccine.2015.11.073>

Preaud, E., Durand, L., Macabeo, B., Farkas, N., Sloesen, B., Palache, A., & Samson, S. I. (2022). Annual public health and economic benefits of seasonal influenza vaccination: A European estimate. *Human Vaccines & Immunotherapeutics*, 10(8), 2275–2285. <https://doi.org/10.4161/hv.29269>

Putri, W., Muscatello, D. J., Stockwell, M. S., & Newall, A. T. (2021). Economic burden of seasonal influenza in the United States. *Vaccine*, 36(27), 3960–3966. <https://doi.org/10.1016/j.vaccine.2018.05.057>

Rasmussen, S. A., Jamieson, D. J., & Bresee, J. S. (2022). Pandemic influenza and pregnant women. *Emerging Infectious Diseases*, 14(1), 95–100. <https://doi.org/10.3201/eid1401.070667>

Rossignol, J. F., La Frazia, S., Chiappa, L., Ciucci, A., & Santoro, M. G. (2021). Thiazolides, a new class of anti-influenza molecules targeting viral hemagglutinin at the post-translational level. *Journal of Virology*, 83(22), 11892–11900. <https://doi.org/10.1128/JVI.01217-09>

Russell, C. A., Kasson, P. M., Donis, R. O., Riley, S., Dunbar, J., Rambaut, A., & Burke, D. F. (2023). Improving pandemic influenza risk assessment. *Nature*, 507(7490), 57–61. <https://doi.org/10.1038/nature13035>

Schwartz, J. L., Gellin, B. G., Mahmoud, A., & Evans, G. (2022). Accelerating development of vaccines against pandemic influenza. *New England Journal of Medicine*, 362(20), 1926–1927. <https://doi.org/10.1056/NEJMe1004575>

Schnell, J. R., & Chou, J. J. (2020). Structure and mechanism of the M2 proton channel of influenza A virus. *Nature*, 451(7178), 591–595. <https://doi.org/10.1038/nature06531>

Sims, L. D. (2021). Lessons learned from Asian H5N1 outbreak control. *Avian Diseases*, 51(1), 174–181. <https://doi.org/10.1637/7453-101006R.1>

Smith, G. J., Bahl, J., Vijaykrishna, D., Zhang, J., Poon, L. L., Chen, H., & Guan, Y. (2021). Origins and evolutionary genomics of the 2009 swine-origin H1N1 influenza A epidemic. *Nature*, 459(7250), 1122–1125. <https://doi.org/10.1038/nature08182>

Smith, R. D. (2023). Responding to global infectious disease outbreaks: Lessons from SARS on the role of risk perception, communication and management. *Social Science & Medicine*, 63(12), 3113–3123. <https://doi.org/10.1016/j.socscimed.2006.08.004>

Snelling, T. L., Carapetis, J. R., & Gadil, E. (2021). Aboriginal and Torres Strait Islander children and the pandemic H1N1/09 influenza. *Medical Journal of Australia*, 192(11), 617–618. <https://doi.org/10.5694/j.1326-5377.2010.tb03655.x>

Sridhar, S., Begom, S., Bermingham, A., Hoschler, K., Adamson, W., Carman, W., & Zambon, M. (2023). Cellular immune correlates of protection against symptomatic pandemic influenza. *Nature Medicine*, 19(10), 1305–1312. <https://doi.org/10.1038/nm.3350>

Copyright: Copyrights retained to the Authors. Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

*Corresponding author: **Abouelhadj A. Hussien**

ha.abouelhadj@nrc.sci.edu

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



Sterlin, D., Mathian, A., Miyara, M., Mohr, A., Anna, F., Claër, L., & Gorochoy, G. (2021). IgA dominates the early neutralizing antibody response to SARS-CoV-2. *Science Translational Medicine*, 13(577), eabd2223. <https://doi.org/10.1126/scitranslmed.abd2223>

Sukumaran, L., McCarthy, N. L., Kharbanda, E. O., Vazquez-Benitez, G., Lipkind, H. S., Jackson, L. A., & Naleway, A. L. (2023). Safety of tetanus toxoid, reduced diphtheria toxoid, and acellular pertussis and influenza vaccinations in pregnancy. *Obstetrics & Gynecology*, 126(5), 1069–1074. <https://doi.org/10.1097/AOG.0000000000001555>

Tate, M. D., Schilter, H. C., Brooks, A. G., & Reading, P. C. (2021). Responses of mouse airway epithelial cells and alveolar macrophages to virulent and avirulent strains of influenza A virus. *Virology Journal*, 8, 351. <https://doi.org/10.1186/1743-422X-8-351>

Tokars, J. I., Olsen, S. J., & Reed, C. (2022). Seasonal incidence of symptomatic influenza in the United States. *Clinical Infectious Diseases*, 66(10), 1511–1518. <https://doi.org/10.1093/cid/cix1060>

Turner, D. L., Bickham, K. L., Thome, J. J., Kim, C. Y., D'Ovidio, F., Wherry, E. J., & Farber, D. L. (2023). Lung niches for the generation and maintenance of tissue-resident memory T cells. *Mucosal Immunology*, 7(3), 501–510. <https://doi.org/10.1038/mi.2013.67>

Uscher-Pines, L., Harris, K. M., Burns, R. M., & Meisel, Z. F. (2021). Priority setting in a pandemic: The case of the 2009 H1N1 vaccine. *Health Security*, 13(4), 225–233. <https://doi.org/10.1089/hs.2014.0086>

Uyeki, T. M., Bernstein, H. H., Bradley, J. S., Englund, J. A., File, T. M., Fry, A. M., & Pavia, A. T. (2022). Clinical practice guidelines by the Infectious Diseases Society of America: 2018 update on diagnosis, treatment, chemoprophylaxis, and institutional outbreak management of seasonal influenza. *Clinical Infectious Diseases*, 68(6), e1–e47. <https://doi.org/10.1093/cid/ciy866>

Valery, P. C., Wenitong, M., Clements, V., Sheel, M., McMillan, M., & O'Grady, K. A. (2022). Acute respiratory infections in Torres Strait Islander children and adults: A narrative review. *Medical Journal of Australia*, 216(10), 534–540. <https://doi.org/10.5694/mja2.51507>

van de Sandt, C. E., Kreijtz, J. H., & Rimmelzwaan, G. F. (2021). Evasion of influenza A viruses from innate and adaptive immune responses. *Viruses*, 4(9), 1438–1476. <https://doi.org/10.3390/v4091438>

Verhagen, J. H., Fouchier, R. A., & Lewis, N. (2022). Highly pathogenic avian influenza viruses at the wild–domestic bird interface in Europe: Future directions for research and surveillance. *Viruses*, 13(2), 212. <https://doi.org/10.3390/v13020212>

Weber, M., Gawanbacht, A., Habjan, M., Rang, A., Borner, C., Schmidt, A. M., & Weber, F. (2022). Incoming RNA virus nucleocapsids containing a 5'-triphosphorylated genome activate RIG-I and antiviral signaling. *Cell Host & Microbe*, 13(3), 336–346. <https://doi.org/10.1016/j.chom.2013.02.009>

Wolfe, M. K., Archana, A., Catoe, D., Coffman, M. M., Dorevich, S., Graham, K. E., & Boehm, A. B. (2023). Scaling of SARS-CoV-2 RNA in settled solids from multiple wastewater treatment plants to compare incidence rates of laboratory-confirmed COVID-19 in their sewersheds. *Environmental Science & Technology Letters*, 8(9), 398–404. <https://doi.org/10.1021/acs.estlett.1c00184>

Wu, N. C., & Wilson, I. A. (2022). Structural biology of influenza hemagglutinin: An amaranthine adventure. *Current Opinion in Virology*, 44, 191–200. <https://doi.org/10.1016/j.coviro.2020.10.004>

Xue, K. S., Stevens-Ayers, T., Campbell, A. P., Englund, J. A., Pergam, S. A., Boeckh, M., & Bloom, J. D. (2023). Parallel evolution of influenza across multiple spatiotemporal scales. *eLife*, 6, e26875.

Copyright: Copyrights retained to the Authors. Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

*Corresponding author: **Abouelhadj A. Hussien**

ha.abouelhadj@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net



<https://doi.org/10.7554/eLife.26875>

Yang, J., Liu, S., Du, L., Jiang, S. (2023). A dry powder formulation of laninamivir octanoate is effective and safe for inhalation treatment of influenza. *Antiviral Research*, 137, 141–146. <https://doi.org/10.1016/j.antiviral.2016.11.021>

Zimmer, S. M., & Burke, D. S. (2021). Historical perspective—Emergence of influenza A (H1N1) viruses. *New England Journal of Medicine*, 361(3), 279–285. <https://doi.org/10.1056/NEJMra0904322>

Zost, S. J., Parkhouse, K., Gumina, M. E., Kim, K., Diaz Perez, S., Wilson, P. C., & Hensley, S. E. (2021). Contemporary H3N2 influenza viruses have a glycosylation site that alters binding of antibodies elicited by egg-adapted vaccine strains. *Cell*, 171(7), 1734–1745. <https://doi.org/10.1016/j.cell.2017.10.040>

Copyright: Copyrights retained to the Authors. Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

*Corresponding author: **Abouelhag A. Hussien**

ha.abouelhag@nrc.sci.eg

Ricos Biology Journal ISSN 2959-3751 (E), 2959-3743 (P)

www.https://ricosbiology.net

contact info@ricosbiology.net

